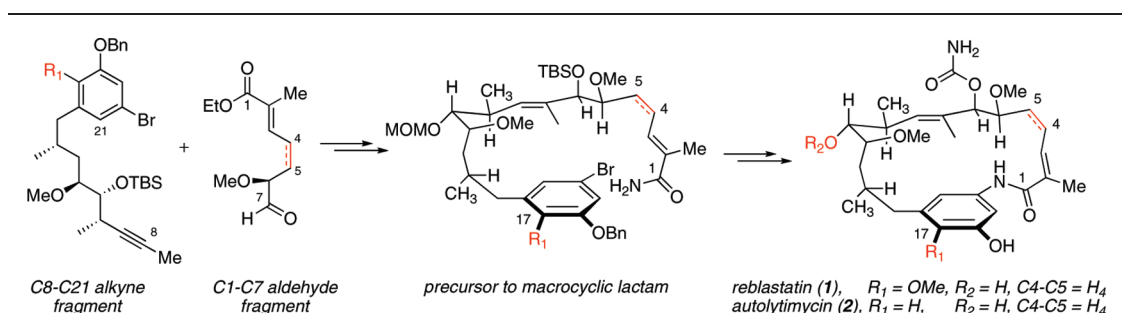# Synthesis of Reblastatin, Autolytimycin, and Non-Benzoquinone Analogues: Potent Inhibitors of Heat Shock Protein 90

Iwona E. Wrona,[†] Alexander Gozman,[‡] Tony Taldone,[‡] Gabriela Chiosis,[‡] and James S. Panek*,[†]

[†]Department of Chemistry and Center for Chemical Methodology and Library Development (CMLD-BU), Metcalf Center for Science and Engineering, 590 Commonwealth Avenue, Boston University, Boston, Massachusetts 02215, and [‡]Department of Medicine and Program in Molecular Pharmacology and Chemistry, Memorial Sloan-Kettering Cancer Center, 1275 York Avenue, New York, New York 10021

panek@bu.edu

Received January 11, 2010

C8-C21 alkyne fragment

C1-C7 aldehyde fragment

precursor to macrocyclic lactam

reblastatin (1), R₁ = OMe, R₂ = H, C4-C5 = H₄
autolytimycin (2), R₁ = H, R₂ = H, C4-C5 = H₄

A full account of an asymmetric synthesis of reblastatin (1) and the first total synthesis of autolytimycin (2) and related structural compounds is described. The syntheses expand the utility of a highly regio- and diastereoselective hydrometalation aldehyde addition sequence to assemble the fully functionalized *ansa* chain of the natural products. Also documented is an intramolecular copper-mediated amidation reaction to close the 19-membered macrolactams. The amidation reaction was also employed for the generation of structural derivatives (6−9) of phenolic ansamycins. Ansamycin natural products and selected structural analogues were evaluated in a competitive binding assay to breast cancer cell lysate and a cytotoxicity assay. Both reblastatin (1) and autolytimycin (2) were shown to bind the heat shock protein 90 with enhanced binding activity (~25 nM) than 17-allylamino-17-demethoxygeldanamycin (17-AAG, 4), a geldanamycin (3) derivative currently under evaluation for treatment of cancer (~100 nM).

## Introduction

Heat shock protein 90 (Hsp90) is a protein chaperone responsible for regulation of proteins in cell signaling, proliferation, and survival processes, including client proteins involved in multiple oncogenic signaling (signal transduction and transcription) pathways.[1,2] As a result, Hsp90 has gained attention as an important therapeutic target for cancer treatment. Hsp90 is effectively inhibited by geldanamycin

(3) and many other benzoquinone ansamycin derivatives,[3a] which bind to the ATP binding site of the N-terminal domain.[3b] The ability of geldanamycin (3)[4] and structurally related agents to affect multiple oncogenic pathways simultaneously is a unique and therapeutically attractive feature of this class of natural products.[5] Despite the cellular potency of geldanamycin (3), the development of this natural product as a clinical agent has been halted due to liver toxicity, insolubility, and cellular instability.[6] The benzoquinone moiety of

(1) For reviews on Hsp90 molecular chaperone see: (a) Wegele, H.; Müller, L.; Buchner, J. *Rev. Physiol., Biochem., Pharmacol.* 2004, *151*, 1. (b) Csermely, P.; Schnaider, T.; Söti, C.; Prohászka, Z.; Nardai, G. *Pharmacol. Ther.* 1998, *79*, 129.

(2) For a short overview of these client proteins see: Goetz, M. P.; Toft, D. O.; Ames, M. M.; Erlichman, C. *Ann. Oncol.* 2003, *14*, 1169 and references cited therein.

(3) (a) Sasaki, K.; Yasuda, H.; Onodera, K. *J. Antibiot.* 1979, *32*, 849. (b) Uehara, Y. *Curr. Cancer Drug Targets* 2003, *3*, 325.

(4) (a) De Boer, C.; Meulman, P. A.; Wnuk, R. J.; Peterson, D. H. *J. Antibiot.* 1970, *23*, 442. (b) Rinehart, K. L.; Sasaki, K.; Slomp, G.; Grostic, M. F.; Olson, E. C. *J. Am. Chem. Soc.* 1970, *92*, 7591.

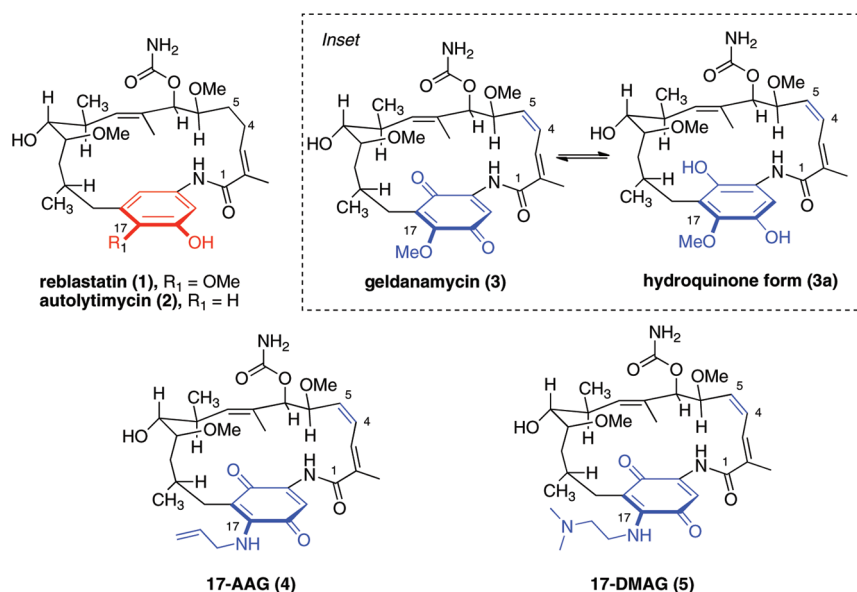(5) Solit, D. B.; Rosen, N. *Curr. Top. Med. Chem.* 2006, *6*, 1205.

**FIGURE 1.** Structure of ansamycin antibiotics.

geldanamycin (**3**) is proposed to be the cause of the observed hepatotoxicity.[7] To stabilize the quinone and increase water solubility of the compound, various 17-aminated semisynthetic derivatives of geldanamycin (**3**) were prepared (Figure 1).[8,9] In turn, less hepatotoxic and more soluble compounds, tanespimycin (**4**) (17-allylamino-17-demethoxygeldanamycin, 17-AAG) and alvespimycin (**5**) (17-(dimethylaminoethylamino)-17-demethoxygeldanamycin, 17-DMAG), are currently in clinical trials for the treatment of cancer.[10,11] However, 17-AAG has been difficult to formulate because of its insolubility characteristics of the quinone and hydroquinone.

Reduction of the quinone to the hydroquinone in ansamycin natural products was shown to increase the binding affinity toward Hsp90.[12,14] A biosynthetic engineering approach has been applied to create nonbenzoquinoid compounds (i.e., phenolic) as Hsp90 inhibitors.[13] Given the close structural resemblance of reblastatin (**1**)[14] and autolytimycin (**2**)[15,16] to geldanamycin (**3**) and its hydroquinone form **3a** (that exhibits enhanced binding affinity over the quinone form; see inset Figure 1), the phenol-containing natural products should bind and inhibit the chaperone activity of Hsp90.[17] The following discussion details the total syntheses of phenol-containing ansamycins **1** and **2**, as well as their structural derivatives **6−9**, and their biological evaluation as effective binders and inhibitors of Hsp90 protein.

Reblastatin (**1**) and autolytimycin (**2**) are polyketide antibiotics that exhibit promising antitumor activity, acting as inhibitors of Hsp90. Reblastatin (**1**) was isolated in 2000 by Takatsu and co-workers during screening experiments intended to identify novel compounds that inhibit phosphorylation of the retinoblastoma protein (Rb).[14] This material was isolated as a minor component from the culture of *Streptomyces hygroscopicus* subsp. hygroscopicus SANK 61995, which also produces the known Hsp90 disruptor geldanamycin (**3**). Like many of the ansamycins, reblastatin's chemical structure is comprised of a 19-membered lactam joined at the meta positions of a phenol ring

(6) (a) Biamonte, M. A.; Van de Water, R.; Arndt, J. W.; Scannevin, R. H.; Perret, D.; Lee, W.-C. *J. Med. Chem.* **2010**, *53*, 3. (b) Thepchatri, P.; Eliseo, T.; Cicero, D. O.; Myles, D.; Snyder, J. P. *J. Am. Chem. Soc.* **2007**, *129*, 3127.

(7) (a) Dikalov, S.; Landmesser, U.; Harrison, D. G. *J. Biol. Chem.* **2002**, *277*, 25480. (b) Tudor, G.; Gutierrez, P.; Aguilera-Gutierrez, A.; Sausville, E. A. *Biochem. Pharmacol.* **2003**, *65*, 1061.

(8) For most recent reviews see: (a) Workman, P. *Trends Mol. Med.* **2004**, *10*, 47. (b) Bagatell, R.; Whitesell, L. *Mol. Cancer Ther.* **2004**, *3*, 1021. (c) Isaacs, J. S.; Xu, W.; Neckers, L. *Cancer Cell* **2003**, *3*, 213. (d) Neckers, L. *Trends Mol. Med.* **2002**, *8*, S55.

(9) (a) Ge, J.; Normant, E.; Porter, J. R.; Ali, J. A.; Dembski, M. S.; Gao, Y.; Georges, A. T.; Grenier, L.; Pak, R. H.; Patterson, J.; Sydor, J. R.; Tibbitts, T. T.; Tong, J. K.; Adams, J.; Palombella, V. J. *J. Med. Chem.* **2006**, *49*, 4606. (b) 8,9-Amido analogues of Geldanamycin: Andrus, M. B.; Wong, Y.; Liu, J.; Beebe, K.; Neckers, L. M. *Tetrahedron Lett.* **2009**, *50*, 6705.

(10) (a) Schulte, T. W.; Neckers, L. M. *Cancer Chemother. Pharmacol.* **1998**, *42*, 273. (b) Banerji, U. *Proc. Am. Assoc. Cancer Ther.* **2003**, *44*, 677. (c) Sausville, E. A.; Tomaszewski, J. E.; Ivy, P. *Curr. Cancer Drug Targets* **2003**, *3*, 377. (d) Jez, J. M; Chen, J. C.-H.; Rastelli, G.; Stroud, R. M.; Santi, D. V. *Chem Biol.* **2003**, *10*, 361. (e) Goetz, M. P.; Toft, D.; Reid, J.; Ames, M.; Stensgard, B.; Safgren, S.; Adjei, A. A.; Sloan, J.; Atherton, P.; Vasile, V.; Salazaar, S.; Adjei, A.; Croghan, G.; Erlichman, C. *J. Clin. Oncol.* **2005**, *23*, 1078.

(11) Kaur, G.; Belotti, D.; Burger, A. M.; Fisher-Nielson, K.; Borsotti, P.; Riccardi, E.; Thillainathan, J.; Hollingshead, M.; Sausville, E. A.; Giavazzi, R. *Clin. Cancer Res.* **2004**, *10*, 4813.

(12) (a) Guo, W.; Reigan, P.; Siegel, D.; Zirrolli, J.; Gustafson, D.; Ross, D. *Cancer Res.* **2005**, *65*, 10006. (b) Maroney, A. C.; Marugan, J. J.; Mezzasalma, T. M.; Barnakow, A. N.; Garrabrant, T. A.; Weaner, L. E.; Jones, W. J.; Barnakova, L. A.; Koblish, H. K.; Todd, M. J.; Masucci, J. A.; Deckman, I. C.; Galemmo, R. A., Jr.; Johnson, D. L. *Biochemistry* **2006**, *45*, 5678. (c) Onuoha, S. C.; Mukund, S. R.; Coulstock, E. T.; Sengerovà, B.; Shaw, J.; McLaughlin, S. H.; Jackson, S. E. *J. Mol. Biol.* **2007**, *372*, 287.

(13) (a) Zhang, M.-Q.; Gaisser, S.; Nur-E-Alam, M.; Sheehan, L. S.; Vousden, W. A.; Gaitatzis, N.; Peck, G.; Coates, N. J.; Moss, S. J.; Radzom, M.; Foster, T. A.; Sheridan, R. M.; Gregory, M. A.; Roe, A. M.; Prodormou, C.; Pearl, L.; Boyd, S. M.; Wilkinson, B.; Martin, C. J. *J. Med. Chem.* **2008**, *51*, 5494. (b) Eichner, S.; Floss, H. G.; Sasse, F.; Kirsching, A. *ChemBioChem* **2009**, *10*, 1801. (c) Roe, S. M.; Prodromou, C.; O'Brien, R.; Ladbury, J. E.; Piper, P. W.; Pearl, L. H. *J. Med. Chem.* **1999**, *42*, 260.
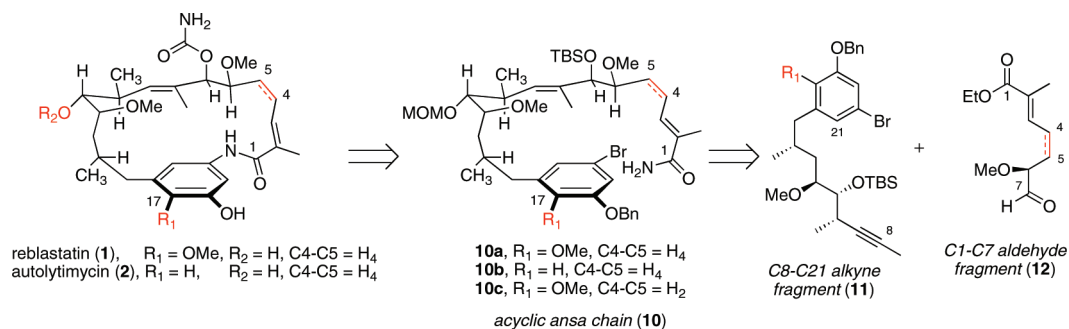
(14) Takatsu, T.; Ohtsuki, M.; Muramatsu, A.; Enokita, R.; Kurakata, S.-I. *J. Antibiot.* **2000**, *53*, 1310.

(15) Li, M. G.; Wu, S. H.; Zhao, L. X.; Zhang, Q.; Li, W. J.; Cui, X. L.; Xu, L. H.; Wu, D. G.; Jiang, C. L. *Chin. Chem. Lett.* **2001**, *12*, 903.

(16) Stead, P.; Latif, S.; Blackaby, A. P.; Sidebottom, P. J.; Deakin, A.; Taylor, N. L.; Life, P.; Spaull, F.; Burrell, F.; Jones, R.; Lewis, J.; Davidson, I.; Mander, T. *J. Antibiot.* **2000**, *53*, 657.

(17) Onodera, H.; Kaneko, M.; Takahashi, Y.; Uochi, Y.; Funahashi, J.; Nakashima, T.; Soga, S.; Suzuki, M.; Ikeda, S.; Yamashita, Y.; Rahayu, E. S.; Kanda, Y.; Ichimura, M. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 1588.

**SCHEME 1.** Retrosynthesis of Reblastatin (**1**), Autolytimycin (**2**), and Structural Derivatives (**6−9**)



reblastatin (**1**),   $R_1$ = OMe, $R_2$ = H, C4-C5 = $H_4$
autolytimycin (**2**),   $R_1$ = H,   $R_2$ = H, C4-C5 = $H_4$

**10a**, $R_1$ = OMe, C4-C5 = $H_4$
**10b**, $R_1$ = H, C4-C5 = $H_4$
**10c**, $R_1$ = OMe, C4-C5 = $H_2$

*acyclic ansa chain* (**10**)

*C8-C21 alkyne fragment* (**11**)

*C1-C7 aldehyde fragment* (**12**)

(Figure 1). The *ansa* chain of this natural product contains six stereogenic centers, two (*E*)-trisubstituted double bonds, and a C7-carbamate functionality. In the initial report, reblastatin (**1**) was shown to inhibit proliferation of cell lines against human histiocytic lymphoma U-937 with an $IC_{50}$ value of 0.43 $\mu$g/mL.[14] Additionally, the natural product was reported to exhibit potent inhibitory activity in the cell-based oncostatin M signaling assay with an $IC_{50}$ value of 0.16 $\mu$M.[16] In 2005, we reported the first total synthesis of reblastatin (**1**), thereby confirming its absolute stereochemistry.[18] The related phenolic natural product autolytimycin (**2**) was isolated in 2001 from a strain of *Streptomyces autolyticus* JX-47 and was shown to exhibit activity in a cell based oncostatin M signaling assay.[15,16] Autolytimycin (**2**) differs structurally from reblastatin (**1**) at the C17 position of the aromatic region, as one has a methoxy group and the other does not. The architectural similarities to geldanamycin (**3**) and other members of the ansamycin family provided incentive to explore these molecules as potential Hsp90 inhibitors. Further, replacement of the *p*-quinone with a phenol ring should attenuate (or possibly remove) the hepatotoxic effects ascribed to the quinone (as in geldanamycin and macbecin) while retaining the over all topology of the ansamycin framework and binding affinity for Hsp90.

### Results and Discussion

**Synthesis of Reblastatin (1), Autolytimycin (2), and Structural Analogues (6−9).** In earlier investigations concerning the chemical synthesis of the ansamycins, macbecin and herbimycin, we took a rather linear approach focusing on the use of crotylsilane reagents in the context of acyclic stereocontrol as reliable means to establish the stereochemical relationships in the *ansa* chain. In that context, we sought to introduce and eventually establish the chiral organosilane reagents bearing C-centered chirality, as carbon nucleophiles that would complement the more established chiral enolate-based bond construction methodology (and aldol surrogates).[19] We had anticipated that these reagents would be

useful for the asymmetric synthesis of polyproprionate derived natural products. Our synthetic strategy to access reblastatin (**1**), autolytimycin (**2**), and structural analogues (**6−9**) explores a more convergent approach (Schemes 1 and 2).[18] The 19-membered macrocycles were envisioned to come from their acyclic frameworks **10** through an *intramolecular* copper(I)-mediated amidation reaction,[20] thereby expanding the scope and utility of the Buchwald aryl amidation methodology.[21] Preparation of the acyclic skeletons **10** would arise from tandem hydrozirconation−transmetalation−nucleophilic addition sequence.[22] The hydrozirconation protocol developed by our laboratories for regioselective preparation of (*E*)-vinyl zirconium species would establish the configuration of the C8−C9 trisubstituted double bond.[23] Introduction of the C7-alcohol would arise from the addition of the in situ derived (*E*)-vinyl metal species (from alkyne **11**) to aldehydes **12**. Structural analogues of the core macrocycles could arise from unsaturation at the C4−C5 positions of aldehyde **12**, functionalization of the aromatic ring, and modification of the secondary C11-alcohol of the *ansa* chain. These minor synthetic changes could provide insight into the necessary chemical modifications needed to produce more potent inhibitors of Hsp90 with less toxicity and higher solubility.

To aid in the synthesis of structural analogues of the phenolic ansamycins, a new approach toward the synthesis of the C8−C21 alkyne fragment was designed (Scheme 2).[24] Advanced alkyne intermediates **11a,b** would now be derived from crotylation of aldehydes **14a,b** with (*Z*)-crotylboron reagent **13**. Our initial synthetic strategy for preparing of these intermediate aldehydes **14a,b** involved crotylation reaction of aldehydes **15a,b** with *syn*-(*E*)-crotylsilane **16**.[18] Following the crotylation with aryl aldehydes **15a,b**, the resulting C15 benzylic oxygen would have to be removed to prepare reblastatin (**1**) or autolytimycin (**2**). To reduce the number of synthetic steps and improve the overall efficiency of this approach, a reductive pyran ring-opening was developed (**17** → **14**; Scheme 2).[24] Tetrahydropyrans **17a,b** can be

(18) (a) Wrona, I. E.; Gabarda, A. E.; Evano, G.; Panek, J. S. *J. Am. Chem. Soc.* **2005**, *127*, 15026. (b) We rationalize that the reversible addition of a second equivalent of $Cp_2ZrHCl$, followed by elimination across the C10−C11 positions may lead to partial epimerization. For mechanistic precedence for this pathway, see: Schwartz, J.; Labinger, J. *Angew. Chem., Int. Ed. Engl.* **1976**, *15*, 333. (c) Wailes, P.; Weigold, H.; Bell, A. P. *J. Organomet. Chem.* **1971**, *27*, 373.

(19) Review on chiral metal enolates: (a) Evans, D. A.; Shaw, J. T. *Ĺactualité Chimique* **2003**, 35–38. (b) Lin, G.-Q.; Li, Y.-M.; Chan, A. S. C. *Principles and Applications of Asymmetric Synthesis*, Wiley & Sons: New York, 2001; pp 135−193. (c) Chiral allyl boranes: Ramachamdran, P. V. *Aldrichimica Acta* **2002**, *35*, 23.

(20) For a recent review on copper-mediated coupling reactions and their applications in natural products synthesis see: Evano, G.; Blanchard, N.; Toumi, M. *Chem. Rev.* **2008**, *108*, 3054.

(21) Klapars, A.; Huang, X.; Buchwald, S. L. *J. Am. Chem. Soc.* **2002**, *124*, 7421.

(22) (a) Wipf, P.; Xu, W. *Tetrahedron Lett.* **1994**, *35*, 5197. (b) Wipf, P.; Jahn, H. *Tetrahedron* **1996**, *52*, 12853. (c) Wipf, P.; Xu, W.; Takahashi, H.; Jahn, H.; Coish, P. D. G. *Pure Appl. Chem.* **1997**, *69*, 639. (d) Wipf, P.; Kendall, C. *Chem.—Eur. J.* **2002**, *8*, 1778.
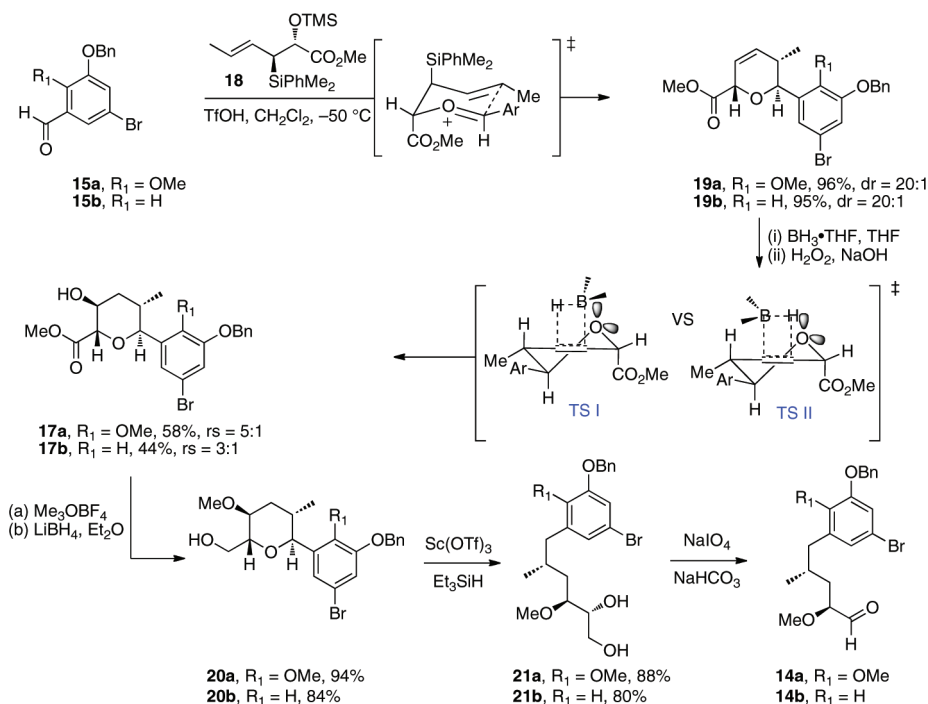
(23) (a) Hu, T.; Panek, J. S. *J. Am. Chem. Soc.* **2002**, *124*, 11368. (b) Panek, J. S.; Hu, T. *J. Org. Chem.* **1997**, *62*, 4912.

(24) (a) Qin, H.-L.; Lowe, J. T.; Panek, J. S. *J. Am. Chem. Soc.* **2007**, *129*, 38. (b) Qin, H.-L.; Panek, J. S. *Org. Lett.* **2008**, *10*, 2477.

**SCHEME 2.** **Retrosynthetic Analysis of Alkyne Fragments 11**
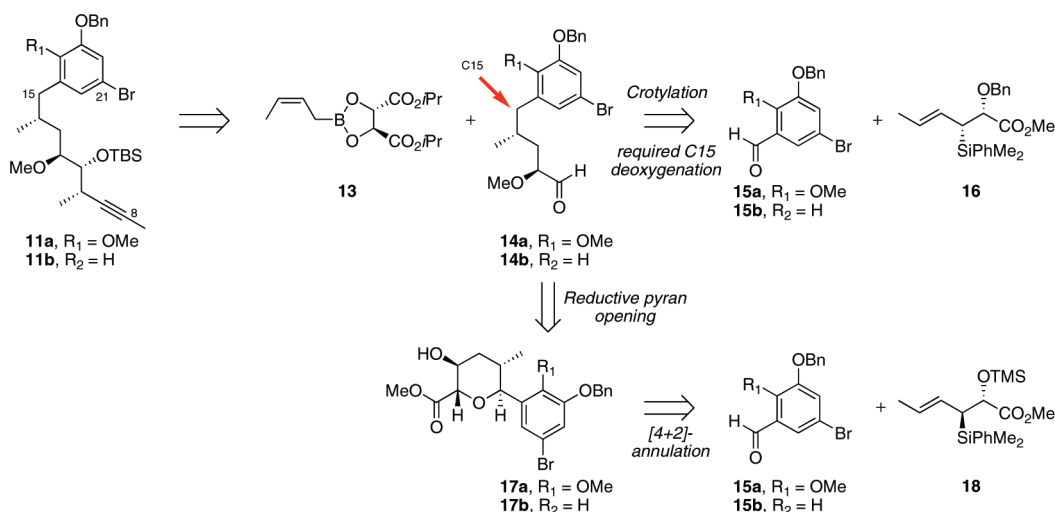


**SCHEME 3.** **Synthesis of the C11−C21 Aldehyde Portions (14a,b) of Alkyne Fragments**



accessed from a Lewis acid-promoted condensation of aromatic aldehydes **15a,b** with *anti*-(*E*)-crotylsilane reagent **18**, allowing for a more direct route toward intermediate aldehydes of type **14**.

**Preparation of the C11−C21 Aldehydes 14.** The synthesis of phenolic ansamycins began with the preparation of the key intermediate aldehydes **14a,b** as depicted in Scheme 3. Exposure of aromatic aldehydes **15a**[25] and **15b**[26] and (*E*)-*anti* crotylsilane **18** to TfOH at −50 °C afforded 2,6-*trans*-

dihydropyrans **19a,b** in greater than 95% yields and excellent selectivities.[27] Hydroboration of the dihydropyran with BH$_3$·THF and subsequent oxidation with alkaline hydrogen peroxide yielded secondary alcohols **17a,b** in moderate yields and regioselectivity; the major isomer is proposed to emerge from an electrostatic and sterically preferred TS I vs TS II.[28] Methylation of alcohols **17a,b** with Meerwein's reagent was followed by reduction of the methyl ester with LiBH$_4$ to give pyran intermediates **20a,b**. Reductive opening of aryl

(25) (a) Prepared in three steps from commercially available 2,3-dihydroxybenzaldehyde; see the Supporting Information for experimental details. (b) For selective benzylation of 2,3-dihydroxy benzaldehyde see: Parker, K. A.; Georges, A. T. *Org. Lett.* **2000**, *2*, 497.

(26) 3-Benzyloxy-5-bromobenzaldehyde was prepared in two steps from commercially available 3,5-dibromophenol. See the Experimental Section for details.

(27) (a) Lowe, J. T.; Panek, J. S. *Org. Lett.* **2005**, *7*, 1529. (b) Huang, H.; Panek, J. S. *Org. Lett.* **2003**, *5*, 1991. (c) Huang, H.; Panek, J. S. *J. Am. Chem. Soc.* **2000**, *122*, 9836.

(28) For related electrophilic additions to dihydropyrans see: (a) Su, Q.; Panek, J. S. *J. Am. Chem. Soc.* **2004**, *126*, 2425–2430. (b) Su, Q.; Panek, J. S. *Angew. Chem., Int. Ed.* **2005**, *44*, 1223.

**SCHEME 4.** Completion of the C9−C21 Aromatic Fragments (11)
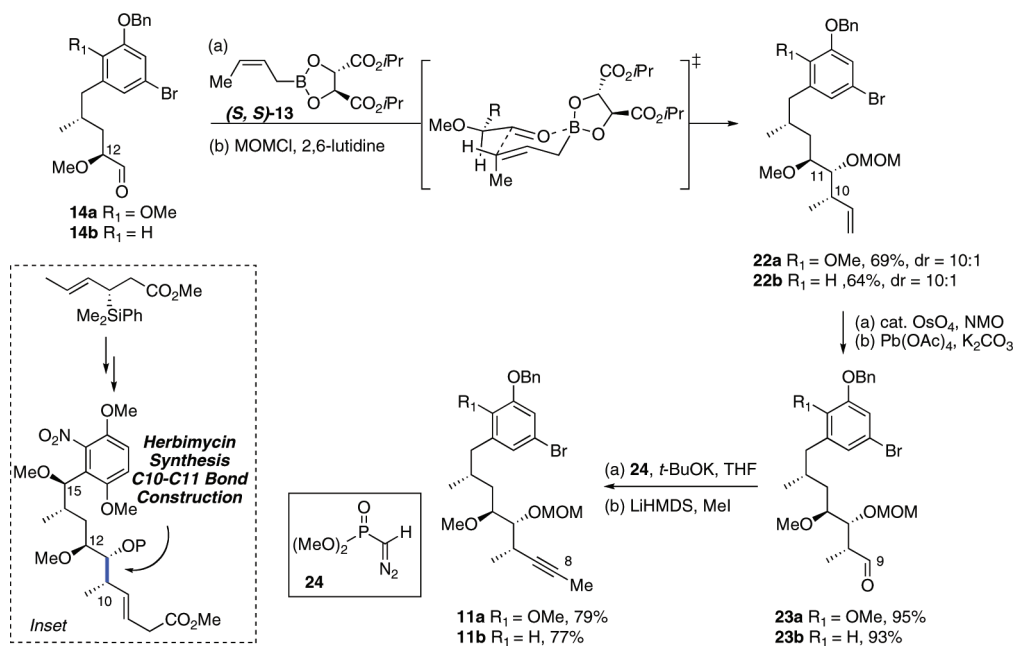


pyranosides **20a,b** was performed in the presence of Sc(OTf)₃ and Et₃SiH to give the C11−C21 aromatic diol fragments **21a,b** in good yields.[24] Notably, the less reactive C17−H aryl pyranoside **20b** required slightly elevated temperature (40 °C vs 25 °C) and longer reaction time for ring-opening. The intermediate diols were then subjected to oxidative cleavage with NaIO₄ providing chiral aldehydes **14a** and **14b** in six steps and 46% and 28% yield, respectively.

Upon completion of the α-methoxy aldehyde fragments **14a,b**, preparation of the necessary C8−C21 subunits required installation of the C10−C11 *syn*-homoallylic alcohols and formation of the C8-methyl substituted alkyne.[18] Although our crotylsilane addition was previously utilized in the synthesis of macbecin I and herbimycin[29] (see inset), for atom economical reasons we elected to use Roush's (*Z*)-crotylboronate reagent (*S,S*)-**13**[30] to simultaneously install the terminal olefin and set the required C10−C11-*syn* stereochemistry. Accordingly, the "matched crotylation" of α-methoxy aldehydes **14a,b** with (*S,S*)-crotylboronate **13** afforded the desired homoallylic alcohols in moderate diastereoselectivity (dr = 10:1) (Scheme 4).[31] The derived homoallylic alcohols were converted to their MOM ethers **22a,b**, followed by dihydroxylation of the terminal alkenes with catalytic osmium tetraoxide. Subsequent oxidative cleavage of the resulting diols exposed the chiral α-methyl aldehydes **23a,b** in 95% and 93% yield, respectively. Treatment with Gilbert−Seyferth reagent **24**[32] provided the terminal alkynes in high yields, and these materials were subjected to methylation by

using LiHMDS and MeI to furnish the desired internal alkynes **11a,b** in 79% and 77% yield, respectively, over the two steps. The fully functionalized C9−C21 aromatic fragments **11a,b** were prepared in 12 steps and 24% and 13% overall yields, respectively, starting from aromatic aldehydes **15a,b**.

**Synthesis of the C1−C7 Aldehyde Fragments 12.** The synthesis of C1−C7 fragments **12a,b** utilized starting materials available from the chiral pool (Scheme 5).[33] The α,β-unsaturated aldehyde **12a** was obtained from readily accessible γ-lactone **26**.[34] Following the reported protocol by Herdeis,[34] L-glutamic acid (**25**) was converted to chiral γ-lactone **26** via a two-step sequence (84% yield). Making use of an analogous strategy employed earlier by Forsyth for the synthesis of the C ring of thyrsiferol,[35] γ-lactone **26** was converted to the (*E*)-α,β-unsaturated ester **27** via a three-step sequence that began with protection of the primary hydroxy group of **26** as its TBS ether, followed by partial reduction of the γ-lactone with DibalH and treatment of the resulting lactol with (carboethoxyethylidene)triphenylphosphorane, thereby providing α,β-unsaturated ester **27** as the (*E*)-isomer in 73% yield (3 steps). Methylation of the secondary alcohol followed by TBS deprotection with dilute HCl and subsequent Swern oxidation[36] of the resulting primary alcohol **28** provided the desired intermediate chiral aldehyde **12a** in 93% yield (3 steps).

The preparation of the C1−C7 aldehyde fragment **12b** required successful installation of the C4−C5 (*Z*)-olefin (Scheme 5). For this reason, we utilized a phosphorus-based olefination approach as a convenient method to install the necessary olefin. Accordingly, 2,3-*O*-isopropylidene-D-glyceraldehyde[37] **29** was treated with (carbomethoxymethylene)-triphenylphosphorane in methanol at 0 °C to afford the desired

(29) For a synthetic review of ansamycin antibiotics see: Wrona, I. E.; Agouridas, V.; Panek, J. S. *C. R. Acad. Sci., Paris* **2008**, *11*, 1483−1522.

(30) Roush, W. R.; Ando, K.; Powers, D. B.; Palkowitz, A. D.; Halterman, R. L. *J. Am. Chem. Soc.* **1990**, *112*, 6339.

(31) (a) Roush, W. R.; Palkowitz, A. D.; Ando, K. *J. Am. Chem. Soc.* **1990**, *112*, 6348. (b) Roush, W. R.; Hoong, L. K.; Palmer, M. A. J.; Park, J. C. *J. Org. Chem.* **1990**, *55*, 4109.

(32) Gilbert, J. C.; Weerasooriya, U. *J. Org. Chem.* **1982**, *47*, 1837. (b) Brown, D. G.; Velthuisen, E. J.; Commerford, J. R.; Brisbois, R. G.; Hoye, T. R. *J. Org. Chem.* **1996**, *61*, 2540.

(33) For examples of chiral pool reagents see: Blaser, H. U. *Chem. Rev.* **1992**, *92*, 935.

(34) Herdeis, C. *Synthesis* **1986**, *232*.

(35) González, I. C.; Forsyth, C. J. *J. Am. Chem. Soc.* **2000**, *122*, 9099.

(36) Omura, K.; Swern, D. *Tetrahedron* **1978**, *34*, 1651.

(37) For preparation of 2,3-*O*-isopropylidene-D-glyceraldehyde from D-mannitol see: Schmid, C.; Bryant, J. D. *Org. Synth.* **1995**, *72*, 6.

**SCHEME 5.** Synthesis of α,β-Unsaturated (12a) and the C1−C7 (*Z,E*)-Diene Fragment (12b)
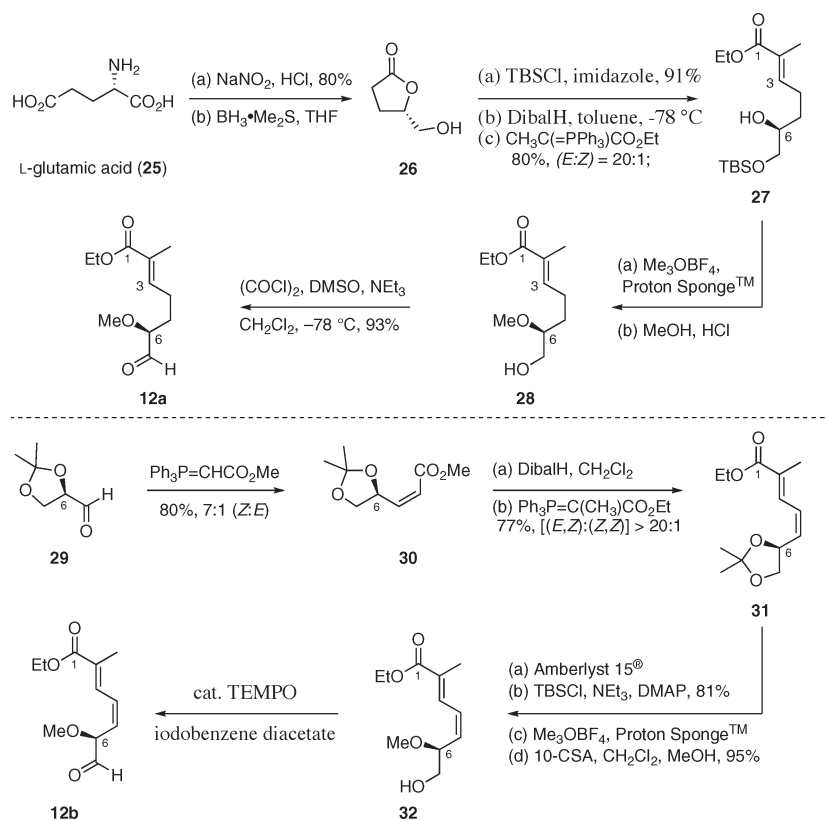


(*Z*)-olefin **30** in 80% yield (*Z*:*E* = 7:1).[38] Reduction of the methyl ester **30** with DibalH and subsequent exposure of the resulting aldehyde to (carboethoxyethylidene)triphenylphosphorane in refluxing benzene proceeded to give (*E,Z*)-diene **31** in 77% yield and excellent selectivity ([(*E,Z*):(*Z,Z*)] = 20:1). Deprotection of the acetonide with Amberlyst 15 yielded an unstable diol that was used without further purification; the primary alcohol was selectively protected as a TBS-ether in 81% over the two steps. Methylation of the secondary alcohol followed by TBS deprotection with 10-camphorsulfonic acid and subsequent TEMPO-catalyzed oxidation[39] of the resulting primary alcohol **32** afforded the desired conjugated *Z,E*-aldehyde **12b** in 94% yield (3 steps).

**Fragment Coupling via Hydrozirconation−Transmetalation−Aldehyde Addition Protocol.** The structural complexity of the advanced coupling partners, methyl-terminated alkynes **11a,b** and α-methoxy aldehydes **12a,b**, represented an interesting synthetic challenge and an opportunity to evaluate a hydro-zirconation−transmetalation−aldehyde addition sequence. In this one-pot process, several transformations occur in a predictable fashion: beginning with the regioselective reduction of the internal alkyne via cis-addition of zirconium and hydride, transmetalation to an organozinc species with retention of the newly formed alkene configuration, and subsequent addition to the aldehyde to afford the desired (*E*)-allylic alcohol. With the execution of a highly convergent and flexible route to access reblastatin (**1**) and autolytimycin

(**2**), we sought to use this synthetic approach to prepare structurally related derivatives to further study the biological activity and structure−activity relationships of this class of ansamycins. This idea could be realized because of the success of the tandem hydrozirconation−transmetalation−nucleophilic addition sequence developed during our synthesis of reblastatin (**1**).[18] The highly regioselective hydrozirconation−transmetalation sequence of alkyne fragments **11a** and **11b** and diastereoselective addition to aldehyde **12a** or **12b** proceeded to give the desired allylic alcohols as single diastereomers in moderate yields (53−70% for **33a**, 40−89% for **33b**, 51−62% for **35**). The stereochemical course of the reaction for the in situ addition of organozinc species to α-methoxy aldehyde is consistent with a Cram-chelate transition state, providing the desired *syn* isomer in 20:1 selectivity.[40] These examples highlight the synthetic utility of the hydrozirconation−transmetalation−nucleophilic addition sequence and ultimately allowed access to a variety of acyclic ansamycins.

The highly regio- and diastereoselective coupling reaction of C1−C7 and C8−C21 fragments afforded the complete acyclic carbon framework of the ansamycins. The synthetic plan for structural analogues is based on the successful assembly of the macrolactam core of reblastatin (**1**).[18] In particular, we wanted to prepare derivatives of reblastatin (**1**) with (*Z,E*)-conjugated diene and modified aromatic functionality at the C17 position. Despite structural studies on complexes of ansamycin derivatives with the ATPase domains of Hsp90, many aspects of their inhibitory mechanism remain unresolved. For instance, it is known that in solution

(38) (a) Mann, J.; Weymouth-Wilson, A. C. *Org. Synth.* **1998**, *75*, 139. (b) Ghosh, A. K.; Leshchenko, S.; Noetzel, M. *J. Org. Chem.* **2004**, *69*, 7822.

(39) De Mico, A.; Margarita, R.; Parlanti, L.; Vescovi, A.; Piancatelli, G. *J. Org. Chem.* **1997**, *62*, 6974.

(40) (a) Reetz, M. T. *Acc. Chem. Res.* **1993**, *26*, 462. (b) Mengel, A.; Reiser, O. *Chem. Rev.* **1999**, *99*, 1191.
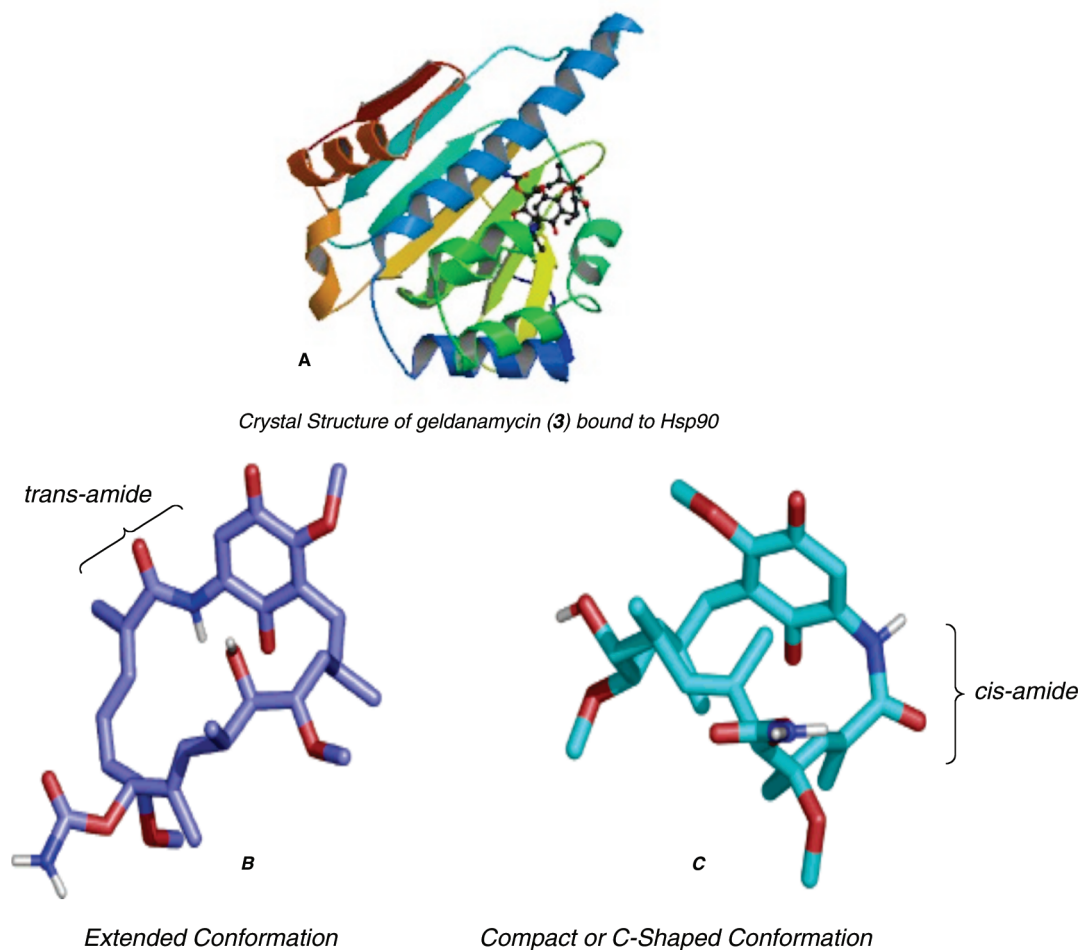
*Crystal Structure of geldanamycin (3) bound to Hsp90*

*trans-amide*



*cis-amide*

**B**

**C**

*Extended Conformation*          *Compact or C-Shaped Conformation*

**FIGURE 2.** (A) Crystal structure of geldanamycin (**3**) bound to Hsp90; solid-state trans-amide (**B**) and protein-bound cis-amide (**C**) conformations of geldanamycin (**3**) (see ref 6b).

geldanamycin (**3**) exists in an extended conformation **A** with the trans geometry about the amide bond.[41] However, it binds Hsp90 in a much more compact conformation (C-shaped) **B** with a cis-amide bond (Figure 2).[6] More specifically, geldanamycin is orientated with the macrocycle and C7 carbamate directed toward the bottom of the binding pocket and the benzoquinone ring directed toward the top of the pocket as it opens to the surface of the binding domain. In contrast to the extended structure adopted by unbound geldanamycin, the protein bound antibiotic is nearly folded over, so that the benzoquinone ring and that of the macrocycle are positioned above and below each other (Figure 2).

Also, the presence of a C17-appendage (excluding a methoxy group) does not significantly affect binding of geldanamycin (**3**) to the protein. In addition to replacing the *p*-quinone ring with a phenol group, our synthetic plan for analogue synthesis was based on examination of the contact points in the crystal structure of geldanamycin (**3**) bound to Hsp90. The analogues incorporated subtle changes within the ansamycin macrocycle to study the effect on binding to Hsp90: incorporation of C4−C5 olefin in reblastatin (**1**), loss

of C17-methoxy group in autolytimycin (**2**), and functionalization of C11-alcohol in reblastatin (**1**). We anticipated that these structural changes could provide additional insight into the necessary structural modifications needed to design more potent ansamycin-derived Hsp90 inhibitors.

The newly formed secondary alcohols were thus converted to the macrocyclization precursors **10a**−**c** (refer to Scheme 1) in a short three-step sequence (Scheme 6). Protection of C7-alcohols **33a**,**b** and **35** as TBS ethers, followed by saponification of the ethyl esters with LiOH and conversion of the resulting acids to the unsaturated amides via a mixed anhydride proceeded in good overall yield (53% for **10a**, 55% for **10b**, 53% for **10c**). It is important to note that the efficiency of the intramolecular amidation reaction was not affected by subtle changes within the *ansa* chain of the ansamycin derivatives and the product macrolactams **34a**,**b** and **36** were accessed in greater than 80% yield for the macrocyclization step. Since our initial publication, the CuI/1,2-diamine protocol was employed in the total synthesis of geldanamycin (**3**)[24b] as well as other 8- to 14-membered lactam-containing natural products.[20,42]

(41) (a) Prodromou, C.; Roe, S. M.; O'Brien, R.; Ladbury, J. E.; Piper, P. W.; Pearl, L. H. *Cell* **1997**, *90*, 65. (b) Stebbins, C. E.; Russo, A. A.; Schneider, C.; Rosen, N.; Hartl, F. U.; Pavletich, N. P. *Cell* **1997**, *89*, 239. (c) Jez, J. M.; Chen, J. C.-H.; Rastelli, G.; Stroud, R. M.; Santi, D. V. *Chem. Biol.* **2003**, *10*, 361.

(42) (a) Hu, T.; Li, C. *Org. Lett.* **2005**, *7*, 2035. (b) Cai, Q.; Zou, B.; Ma, D. *Angew. Chem., Int. Ed.* **2006**, *45*, 1276. (c) Toumi, M.; Couty, F.; Evano, G. *J. Org. Chem.* **2007**, *72*, 9003. (d) Toumi, M.; Couty, F.; Evano, G. *Angew. Chem., Int. Ed.* **2007**, *46*, 572.

**SCHEME 6.** Synthesis of the Macrocyclic Cores of Phenolic Derivatives[a]
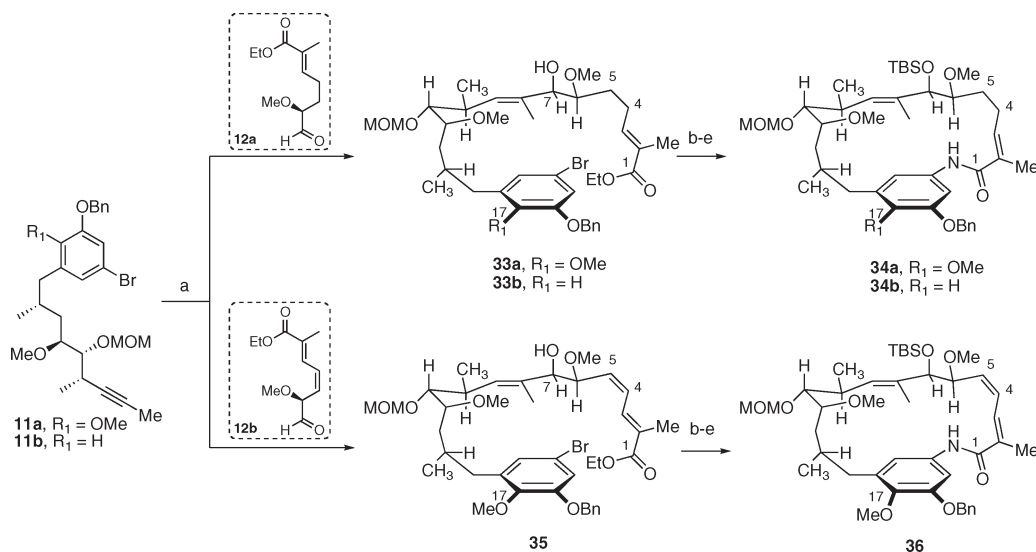


[a]Reagents and conditions: (a) (i) 2 equiv of Cp$_2$ZrHCl, **11a** or **11b**, toluene, 50 °C, (ii) ZnMe$_2$, toluene, −65 °C, (iii) **12a** or **12b**, 0 °C, 53−70% for **33a**, 40−89% for **33b**, 51−62% for **35**, dr = 20:1; (b) TBSOTf, 2,6-lutidine, CH$_2$Cl$_2$, 0 °C; (c) LiOH, THF/MeOH/H$_2$O; (d) (i) (CH$_3$)$_2$CHCH$_2$OCOCl, NEt$_3$, CH$_2$Cl$_2$, −20 °C, (ii) NH$_3$ (*l*); (e) CuI, *N,N′*-dimethylethylenediamine, K$_2$CO$_3$, toluene, 100 °C, 36 h, >80% for **34a**, 80% yield for **34b**, 82% yield for **36**.

Conversion of the desired macrocyclic cores **34a,b** and **36** en route to the ansamycin derivatives is depicted in Scheme 7. Toward that end, a three-step sequence was employed to convert the C7-silyl ethers to the required carbamate functionality. Deprotection of the initial silyl ethers **34a,b** and **36** was achieved by using buffered HF·pyridine. The resulting C7-secondary alcohols were converted to their carbamates using reaction conditions reported by Kocovsky[43] and provided the desired materials in greater than 80% yield. Initial attempts to selectively remove the MOM ether under mild conditions[44] resulted in decomposition of the macrolactam. Ultimately, deprotection of the MOM and benzyl ethers was accomplished by using AlCl$_3$ and anisole[45] to reveal reblastatin (**1**), autolytimycin (**2**), and diene derivative **7** in 75%, 61%, and 63% yield, respectively. Similarly, exposure of the intermediate carbamate derived from **34a** to BCl$_3$ at low temperatures provided reblastatin (**1**) in 53% isolated yield, with recovered MOM-protected reblastatin **6** in 15% yield. Both synthetic reblastatin (**1**) and autolytimycin (**2**) exhibited physical, spectroscopic, and spectrometric characteristics ($^1$H, $^{13}$C NMR, IR, [α]$_D$, and HRMS) identical with those reported for the natural products.[46]

Employing a similar strategy, we were able to functionalize the C11 secondary alcohols of the ansamycin derivatives as carbamates. Sequential treatment of macrolactams **34a** and **36** with a solution of MgBr$_2$ and HF·Pyr afforded the respective C7,C11-diols which were treated with excess trichloroisocyanate reagent to afford the bis-carbamates. The resulting C7,C11 bis-carbamate derivatives bearing benzyl ether-protected C18 phenolic oxygens were then treated with AlCl$_3$ to provide the desired phenolic ansamycins **8** and **9** in 82% and 50% yields, respectively, over the four steps.

The convergent route to reblastatin (**1**), autolytimycin (**2**), and related structures (**6**−**9**) required an average of 23 steps with overall yields ranging between 2.79% and 5.67%, which underscores the critical role that chemical synthesis plays in the production of new chemical entities, and in the present case, compounds possessing enhanced binding affinity for Hsp90.

**Biological Evaluation for Hsp90 Inhibition.** The synthesis of natural product-like libraries (and small molecule collections) comprised of sophisticated macrocycles rich in topological or stereochemical variation is an underdeveloped field with enormous potential in biomedical research.[47] Many molecules in nature are produced through convergent biosynthesis to afford natural products (often called hybrids) whose biological properties come about through their enhanced interactions with proteins.[48]

Having designed and executed a convergent and flexible approach to access the macrocyclic lactams reblastatin (**1**) and autolytimycin (**2**) and selected related structures, we sought to explore their effectiveness as inhibitors of Hsp90. In that regard, reblastatin (**1**), autolytomycin (**2**), and four structural derivatives **6**−**9** were evaluated in a competitive binding assay from MDA-MB-468 breast cancer lysate.[18]

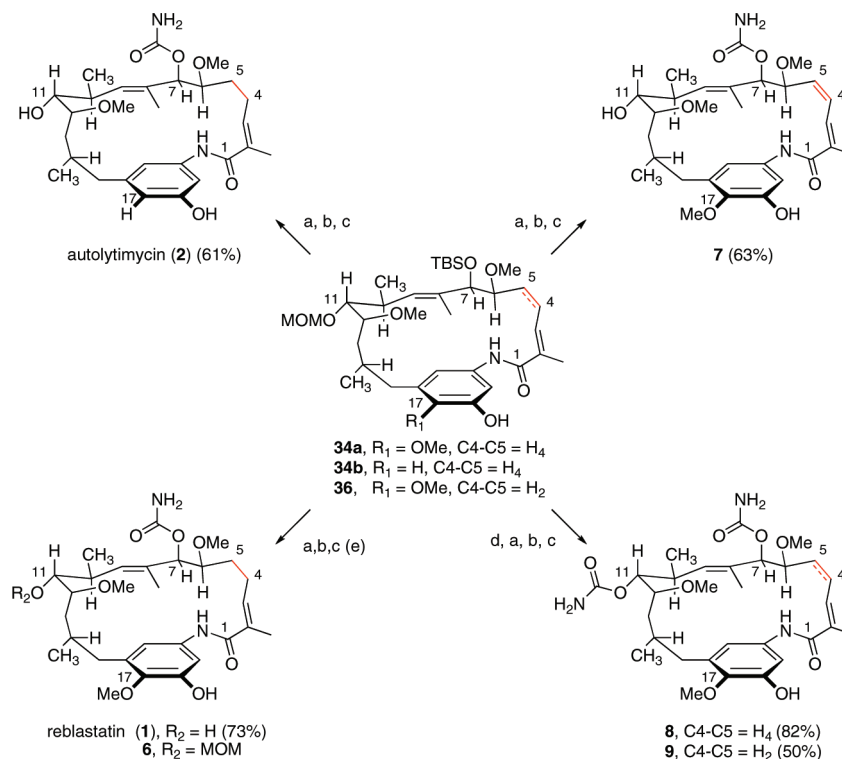(43) Kocovsky, P. *Tetrahedron Lett*. **1986**, *27*, 5521.

(44) Attempts to deprotect the C11 MOM-ether with the following procedures failed: (a) Catecholborane: Boeckman, R. K., Jr.; Potenza, J. C. *Tetrahedron Lett*. **1985**, *26*, 1411. (b) MgBr$_2$: Kim, S.; Kee, I. S.; Park, Y. H.; Park, J. H. *Synlett* **1991**, 183. (c) CBr$_4$/*i*-PrOH: Lee, A. S.-Y.; Hu, Y.-J.; Chu, S.-F. *Tetrahedron* **2001**, *57*, 2121. (d) TMSBr: Hanessian, S.; Delorme, D.; Dufresne, Y. *Tetrahedron Lett*. **1984**, *25*, 2515. (e) BCl$_3$·SMe$_2$: Congreve, M. S.; Davison, E. C.; Fuhry, M. A. M.; Holmes, A. B.; Payne, A. N.; Robinson, R. A.; Ward, S. E. *Synlett* **1993**, 663. (f) Me$_2$BBr: Guindon, Y.; Morton, H. E.; Yoakim, C. *Tetrahedron Lett*. **1983**, *24*, 3969. (g) AcCl/MeOH/Et$_2$O: Tius, M. A.; Gomex-Galeno, J.; Gu, X. Q.; Zaidi, J. H. *J. Am. Chem. Soc*. **1991**, *113*, 5775.

(45) Akiyama, T.; Hirofuji, H.; Ozaki, S. *Tetrahedron Lett*. **1991**, *32*, 1321.

(46) See the Supporting Information for comparison of spectral data ($^1$H NMR and $^{13}$C NMR).

(47) Recent examples: (a) Beeler, A. B.; Acquilano, D. E.; Su, Q.; Yan, F.; Roth, B. L.; Panek, J. S.; Porco, J. A., Jr. *J. Comb. Chem*. **2005**, *7*, 673–681. (b) Dandapani, S.; Beeler, A. B.; Lan, P.; Beischel, S.; Abbas, A.; Roth, R. B.; Porco, J. A., Jr.; Panek, J. S. *J. Org. Chem*. **2006**, *71*, 8934.

(48) Tietze, L. F.; Bell, H. P.; Chandrasekhar, S. *Angew. Chem., Int. Ed*. **2003**, *42*, 3996.

SCHEME 7. Synthesis of Reblastatin (**1**), Autolytimycin (**2**), and Structural Analogues (**6**−**9**)[a]



**34a**, R$_1$ = OMe, C4-C5 = H$_4$
**34b**, R$_1$ = H, C4-C5 = H$_4$
**36**, R$_1$ = OMe, C4-C5 = H$_2$

autolytimycin (**2**) (61%)

**7** (63%)

reblastatin (**1**), R$_2$ = H (73%)
**6**, R$_2$ = MOM

**8**, C4-C5 = H$_4$ (82%)
**9**, C4-C5 = H$_2$ (50%)

[a]Reagents and conditions: (a) HF·pyridine, pyridine, THF; (b) (i) Cl$_3$CCONCO, CH$_2$Cl$_2$, (ii) MeOH, K$_2$CO$_3$; (c) AlCl$_3$, anisole, CH$_2$Cl$_2$, −78 °C to rt; (d) MgBr$_2$, EtSH, Et$_2$O, rt; (e) BCl$_3$, CH$_2$Cl$_2$, −78 °C, 15% for **6**.

We anticipated that reblastatin (**1**), bearing a phenolic hydroxyl, would bind to the molecule chaperone Hsp90 in a similar manner to that of the natural product geldanamycin (**3**).[17] Additionally, synthetic derivatives devoid of the benzoquinone moiety and C-17 methoxy group could potentially provide new agents with improved toxicity profiles over geldanamycin (**3**), 17-AAG (**4**), and 17-DMAG (**5**). All evidence to date confirms that the reduction of the quinone moiety in geldanamycin derivatives increases the drug's affinity for Hsp90.[49] Growing experimental and clinical evidence of the potential of these phenolic molecules as anticancer agents supports the need to develop chemical syntheses of this class of natural products like reblastatin (**1**) and its structurally related agents.[6]

After completion of the total synthesis of reblastatin (**1**), the natural product and its select analogues **6**, **37**, and **38** (Figure 3) were evaluated for their binding affinity for Hsp90. Reblastatin (**1**) was shown to exhibit 4-fold enhanced nanomolar activity (7.34 nM) over geldanamycin **3** (34−42 nM) in a competitive binding assay to Hsp90 derived from MDA-MB-468 breast caner cell lysate (Figure 3).[50] Further, the MOM-ether derivative **6** also displayed Hsp90 inhibition (70.3 nM), whereas removal of C7-urethane **38** or an acyclic framework derivative **37** were ineffective as Hsp90 inhibitors; findings are consistent with previous SAR studies on geldanamycin (**3**).[8,9]

The structure of the geldanamycin-binding domain of Hsp90 reveals a pronounced pocket that is highly conserved

across species. Geldanamycin binds inside this pocket, adopting a compact structure similar to that of a polypeptide chain in a turn conformation. In contrast to the extended structure adopted by unbound geldanamycin, the protein bound antibiotic is nearly folded over, so that the benzoquinone ring and that of the macrocycle are positioned above and below each other (Figure 2).[13c] The preliminary competitive binding assays of reblastatin (**1**) and related structural analogues revealed several important structural interactions between Hsp90 and phenol-containing ansamycins. The removal of the C21-hydroxyl (of the reduced hydroquinone of geldanamycin) and its associated polar contact with the protein resulted in higher binding affinity for the molecular chaperone. Suggesting that binding to Hsp90 is not dependent on the presence of the quinone moiety and/or loss of C4−C5 olefin (e.g. geldanamycin), thus inhibition of the molecular chaperone can be extended to phenol-containing ansamycins.[17]

Having achieved a successful total synthesis of autolytimycin (**2**) and four structurally similar derivatives (**7**−**9**), we were positioned to examine these structures as inhibitors of the bioactivity of Hsp90 in cancer cells. Along with reblastatin (**1**), the ansamycin derivatives **2** and **7**−**9** were evaluated for cytotoxicity in two acute myeloid leukemia cell lines and for binding affinity to Hsp90. The Kasumi-1 and MOLM-13 cell growth inhibition (IC$_{50}$) assays and the apparent binding affinities (EC$_{50}$) to recombinant human Hsp90 were measured by using previously reported protocols (Figure 4).[51]

(49) Neckers, L. *Trends Mol. Med.* **2002**, *8*, S55.
(50) See the Supporting Information for details.

(51) (a) For competition assays see: Moulick, K.; Clement, C. C.; Aguirre, J.; Kim, J.; Kang, Y.; Felts, S.; Chiosis, G. *Bioorg. Med. Chem. Lett.* **2006**, *16*, 4515. (b) For growth assays see: O'Brien, J.; Wilson, I.; Orton, T.; Pognan, F. *Eur. J. Biochem.* **2000**, *267*, 5421.
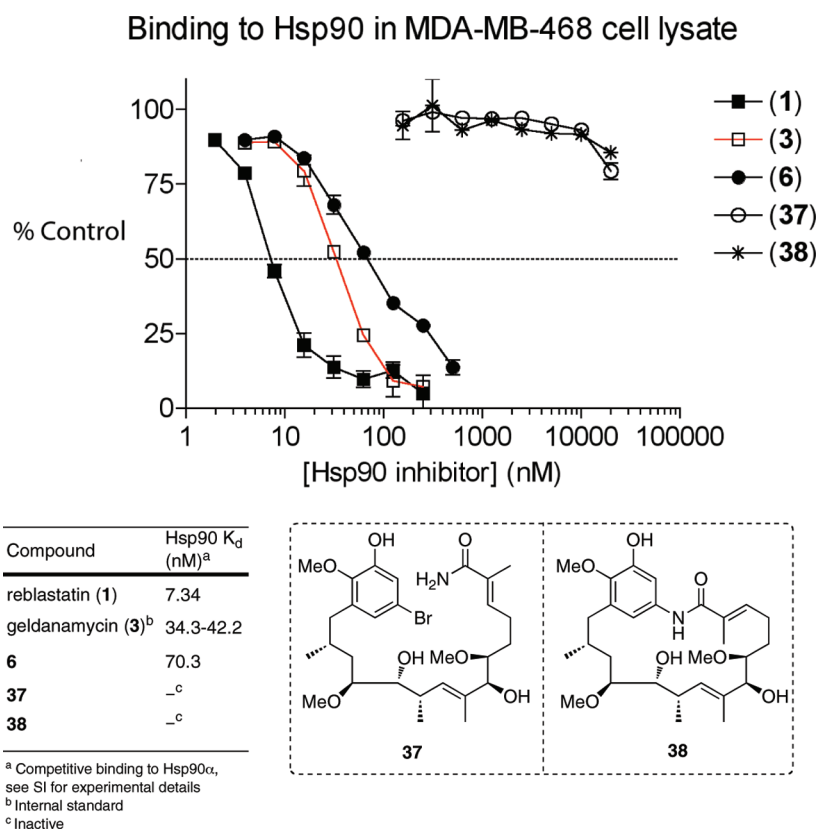
## Binding to Hsp90 in MDA-MB-468 cell lysate



| Compound | Hsp90 $K_d$ (nM)[a] |
|---|---|
| reblastatin (**1**) | 7.34 |
| geldanamycin (**3**)[b] | 34.3-42.2 |
| **6** | 70.3 |
| **37** | –[c] |
| **38** | –[c] |

[a] Competitive binding to Hsp90α, see SI for experimental details
[b] Internal standard
[c] Inactive

**FIGURE 3.** Competitive binding assay for reblastatin (**1**) and structural analogues **6**, **37**, and **38** to Hsp90 obtained from MDA-MB-468 breast cancer lysate. Reblastatin (**1**) binds to Hsp90 with slightly better nanomolar activity (7.34 nM) than geldanamycin (**3**, 34–42 nM). Derivative **6** also showed good inhibitory activity (70.3 nM). Compounds **37** and **38** were inactive in the binding assay.

The results for the phenolic analogues are shown in Table 1. The affinity of Hsp90 to reblastatin **1** (26 nM) and autolytimycin **2** (36 nM) natural products is about 4-fold higher than that for the corresponding control, 17-AAG **4** (110 nM). Interestingly, the C4−C5 olefin containing derivative **7** showed a $K_d$ value (86 nM) similar to that of 17-AAG (**4**), while the two bis-urethane analogues (**8** and **9**) exhibited weak binding affinity (about 10-fold lower) altogether. The natural products **1** and **2** were both more potent (IC$_{50}$ 280 nM) at inhibition of Kasumi-1 cell growth than 17-AAG **4** (IC$_{50}$ 480 nM). Similar to the Hsp90 binding data, the C4−C5 olefin analogue **7** was slightly less potent (IC$_{50}$ 760 nM) in the Kasumi-1 assay. Interestingly, the phenolic ansamycins were less potent against the MOLM-13 cell line with reblastatin (**1**) being 3-fold less active ($K_d \approx 270$ nM) than 17-AAG **4** ($K_d \approx 90$ nM), while autolytimycin (**2**) was about 6-fold less active ($K_d \approx 510$ nM). The binding assay of the phenolic ansamycins revealed that modification to the aromatic moiety and the C4−C5 olefin are well tolerated for Hsp90 binding affinity, while modifications at the C-11 alcohol (addition of carbamate functional group) resulted in substantial loss of activity. The analogues were also screened against two acute myeloid leukemia cell lines (Kasumi-1 and MOLM-13). Reblastatin (**1**) and autolytimycin (**2**) were more potent than 17-AAG (**4**) in inhibiting cell growth in Kasumi-1, but were 3- and 5-fold, respectively, less active in the MOLM-13 cell line. The binding affinity observed for these natural product derivatives confirms that in vitro binding to the molecular target

Hsp90 is necessary for activity but not sufficient for cytotoxicity.[52]

### Conclusions

In summary, an effective synthetic route for a second-generation synthesis of reblastatin (**1**), the first total synthesis of autolytimycin (**2**), and structurally related ansamycin analogues (**6−9**) were developed. The natural products were prepared in highly enantioenriched form, with a longest linear sequence of 26 steps. A regio- and diastereoselective hydrometalation−transmetalation−aldehyde addition sequence was utilized to assemble the fully functionalized *ansa* frameworks of the natural products and derivatives. Additionally, the copper-mediated amidation reaction tolerated subtle changes within the *ansa* chain and aromatic moiety to produce structurally similar macrocycles in high yields. The synthetic natural products were tested for Hsp90 binding affinities and cytotoxicity assays. Although the natural products reblastatin (**1**) and autolytimycin (**2**) were shown to bind the Hsp90 protein with slightly better affinity (4-fold over 17-AAG **4**), the synthetic derivatives were less potent in the growth inhibition assays. Further investigation of these synthetic derivatives is needed to determine the in vitro efficacy and pharmacological profiles of these compounds over 17-AAG (**4**).

(52) Tian, Z.-Q.; Liu, Y.; Zhang, D.; Wang, Z.; Dong, S. D.; Carreras, C. W.; Zhou, Y.; Rastelli, G.; Santi, D. V.; Myles, D. C. *Bioorg. Med. Chem.* **2004**, *12*, 5317.
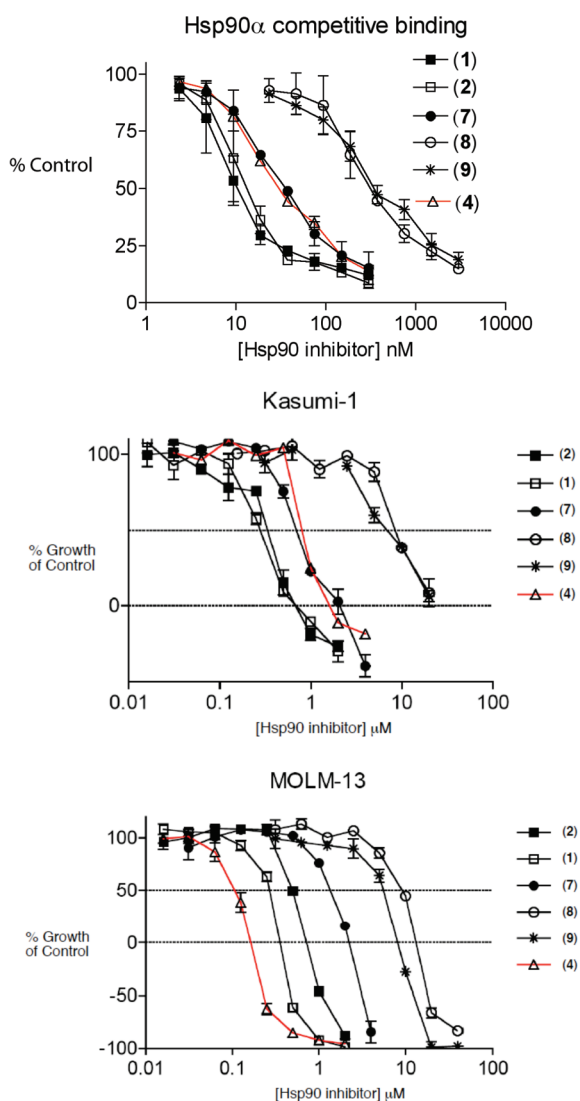
## Hsp90α competitive binding



## Kasumi-1



## MOLM-13



**FIGURE 4.** Cytotoxicity and Hsp90 binding affinity of reblastatin (**1**), autolytimycin (**2**), 17-AAG (**4**), and analogues (**7−9**).

**TABLE 1.** Cytotoxicity and Hsp90 Binding Affinity of Reblastatin (**1**), Autolytimycin (**2**), 17-AAG (**4**), and Analogues (**7−9**)

| compd | Hsp90 $K_d$ (nM)[a] | Kasumi-1 IC$_{50}$ (nM)[b] | MOLM-13 IC$_{50}$ (nM)[b] |
|---|---|---|---|
| reblastatin (**1**) | 26 | 290 | 270 |
| autolytimcyin (**2**) | 36 | 280 | 510 |
| **7** | 84 | 760 | 1550 |
| **8** | 1475 | 7280 | 5930 |
| **9** | 1265 | 9030 | 9590 |
| 17-AAG (**4**)[c] | 110 | 480 | 90 |

[a]Competitive binding to Hsp90α, see the SI for experimental details. [b]See the SI for experimental details. [c]Internal standard.

## Experimental Section

The following experimental information is representative and describes the complete details of the convergent synthesis of autolytimycin (**2**).

**1-(Benzyloxy)-3,5-dibromobenzene.** To a solution of 3,5-dibromophenol (17.3 g, 68.7 mmol) in acetone (140 mL) at room temperature was added potassium carbonate (14.2 g, 103 mmol) followed by benzyl bromide (8.98 mL, 75.5 mmol). The reaction

was allowed to run at room temperature for 16 h. The heterogeneous mixture was filtered over Celite, rinsed with ethyl acetate, and concentrated under reduced pressure to give a yellow oil. Purification by flash chromatography (silica, 10% EtOAc/hexanes) affords bis-bromo phenol as a colorless oil (23.0 g, 67.2 mmol, 97.8%). $^1$H NMR (CDCl$_3$, 400 MHz) $\delta$ 7.57−7.50 (m, 5H), 7.43 (dt, $J$ = 1.6, 3.4 Hz, 1H), 7.24 (d, $J$ = 1.6 Hz, 2H); $^{13}$C NMR (CDCl$_3$, 100 MHz) $\delta$ 159.8, 135.7, 128.7, 128.3, 127.5, 126.6, 123.1, 117.2, 70.5; IR (neat) $\nu_{max}$ 3080, 3033, 2933, 2871, 1583, 1436, 1256, 1024, 830, 746, 696 cm$^{-1}$; HRMS (CI, NH$_3$) $m/z$ calcd for C$_{13}$H$_{10}$Br$_2$O [M + 23]$^+$ 362.8996, found 362.8986.

**3-(Benzyloxy)-5-bromobenzaldehyde (15b).** To a solution of 1-(benzyloxy)-3,5-dibromobenzene (12.2 g, 35.7 mmol) in tetrahydrofuran (360 mL) at −78 °C was added *n*-BuLi in hexane (2.50 M, 15.7 mL) dropwise. The reaction mixture was stirred for 30 min and *N*,*N*-dimethylformamide (4.14 mL, 53.5 mmol) was added in one portion. The reaction mixture was warmed to room temperature and stirred for an additional 30 min. The reaction solution was poured into a vigorously stirring solution of 10% aq KH$_2$PO$_4$ and diethyl ether (310 mL:190 mL). The biphasic mixture was stirred for an additional 10 min. The organic layer was separated and the aqueous solution was extracted with diethyl ether (2×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. Purification by flash chromatography (silica, 5% EtOAc/hexanes) gives aldehyde **15b** as a white solid (9.0 g, 30.9 mmol, 86.6%). Mp 52 °C; $^1$H NMR (CDCl$_3$, 400 MHz) $\delta$ 9.88 (s, 1H), 7.58 (t, $J$ = 1.6 Hz, 1H), 7.42−7.33 (m, 7H), 5.09 (s, 2H); $^{13}$C NMR (CDCl$_3$, 100 MHz) $\delta$ 190.5, 159.8, 138.6, 135.6, 128.7, 128.4, 127.5, 125.9, 124.6, 123.5, 113.0, 70.5; IR (neat) $\nu_{max}$ 3068, 3033, 2837, 2727, 1702, 1570, 1270, 1028, 847, 696 cm$^{-1}$; HRMS (CI, NH$_3$) $m/z$ calcd for C$_{14}$H$_{11}$BrO$_2$ [M + 23]$^+$ 312.9840, found 312.9851.

**(2S,5S,6S)-Methyl 6-(3-(Benzyloxy)-5-bromophenyl)-5-methyl-5,6-dihydro-2H-pyran-2-carboxylate (19b).** To a solution of benzaldehyde **15b** (9.00 g, 30.9 mmol) and (*E*)-crotylsilane **18** (12.5 g, 35.5 mmol) in methylene chloride (600 mL) at −50 °C was added trifluoromethanesulfonic acid (2.74 mL, 30.9 mmol) dropwise. The reaction mixture was allowed to stir for 12 h at −50 °C before it was quenched by addition of a saturated solution of sodium bicarbonate. The aqueous phase was extracted with methylene chloride (2×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. Purification by flash chromatography (silica, 10% EtOAc/hexanes) yields dihydropyran **19b** as a clear oil (12.0 g, 28.8 mmol, 93.0%). [α]$^{20}$$_D$ −71.5 (*c* 1.0, CHCl$_3$); $^1$H NMR (CDCl$_3$, 400 MHz) $\delta$ 7.41−7.30 (m, 5H), 7.16 (t, $J$ = 1.6 Hz, 1H), 7.07 (t, $J$ = 2.0 Hz, 1H), 6.95 (dt, $J$ = 1.2, 2.4 Hz, 1H), 5.89 (tq, $J$ = 2.0, 2.8, 9.8, 24.6 Hz, 2H), 5.03 (s, 2H), 4.86 (q, $J$ = 2.8, 6.0 Hz, 1H), 4.32 (d, $J$ = 9.6 Hz, 1H), 3.75 (s, 3H), 2.39 (m, 1H), 0.81 (d, $J$ = 7.2 Hz, 3H); $^{13}$C NMR (CDCl$_3$, 100 MHz) $\delta$ 171.2, 159.4, 143.3, 136.3, 133.6, 128.6, 128.1, 127.5, 123.3, 122.7, 122.4, 117.7, 113.3, 79.3, 73.1, 70.3, 52.1, 34.7, 16.4; IR (neat) $\nu_{max}$ 3037, 2955, 2875, 1753, 1570, 1443, 1272, 1148, 1109, 782 cm$^{-1}$; HRMS (CI, NH$_3$) $m/z$ calcd for C$_{21}$H$_{21}$BrO$_4$ [M + 23]$^+$ 439.0521, found 439.0535.

**(2S,3S,5S,6S)-Methyl 6-(3-(Benzyloxy)-5-bromophenyl)-3-hydroxy-5-methyltetrahydro-2H-pyran-2-carboxylate (17b).** To a solution of dihydropyran **19b** (6.00 g, 14.4 mmol) in tetrahydrofuran (150 mL) at 0 °C was added BH$_3$·THF complex in tetrahydrofuran (1.11 M, 22.0 mL) dropwise. The reaction mixture was allowed to warm to room temperature on its own and stirred for an additional 1 h at room temperature (about 3 h). The reaction was cooled to 0 °C and quenched by sequential addition of sodium hydroxide in water (2.00 M, 35.9 mL) and 30% H$_2$O$_2$ (14.7 mL). The quenched reaction was stirred for

$^1/_2$ h at 0 °C and for $^1/_2$ h at room temperature. The mixture was diluted with water and the aqueous layer was extracted with diethyl ether (5×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. Purification by flash chromatography (silica, 30% EtOAc/hexanes) provides alcohol **17b** as a white foam (3.00 g, 28.8 mmol, 47.0%). $[\alpha]^{20}{}_D$ −7.5 (*c* 1.0, CHCl$_3$); $^1$H NMR (CDCl$_3$, 400 MHz) δ 7.41−7.30 (m, 5H), 7.14 (t, *J* = 1.4 Hz, 1H), 7.06 (t, *J* = 2.2 Hz, 1H), 6.92 (dd, *J* = 1.4, 2.2 Hz, 1H), 5.03 (s, 2H), 4.49 (t, *J* = 1.8 Hz, 1H), 4.33 (m, 2H), 3.79 (s, 3H), 2.20 (d, *J* = 7.2 Hz, 1H), 2.10 (m, 1H), 1.94 (ddt, *J* = 1.6, 3.6, 14.4 Hz, 1H), 1.50 (ddd, *J* = 2.8, 3.6, 12.2, 14.4 Hz, 1H) 0.70 (d, *J* = 6.4 Hz, 3H); $^{13}$C NMR (CDCl$_3$, 100 MHz) δ 170.4, 159.3, 143.2, 136.2, 128.5, 128.1, 127.5, 123.1, 122.6, 117.4, 113.4, 81.5, 78.3, 70.2, 65.8, 52.2, 35.8, 29.6, 17.5; IR (neat) $\nu_{max}$ 3412, 2953, 2928, 2872, 1747, 1570, 1441, 1270, 1136, 1057, 696 cm$^{-1}$; HRMS (CI, NH$_3$) *m/z* calcd for C$_{21}$H$_{23}$BrO$_5$ [M + 23]$^+$ 457.0627, found 457.0672.

**(2S,3S,5S,6S)-Methyl 6-(3-(Benzyloxy)-5-bromophenyl)-3-methoxy-5-methyltetrahydro-2*H*-pyran-2-carboxylate.** To a solution of alcohol **17b** (5.70 g, 13.1 mmol) in methylene chloride (6.50 mL) at room temperature was added sequentially 4 Å molecular sieves (8.50 g), *N,N,N′,N′*-tetramethyl-1,8-naphthalenediamine (8.42 g, 39.3 mmol), and trimethyloxonium tetrafluoroborate (4.84 g, 32.7 mmol). The reaction mixture was allowed to stir for 16 h at room temperature and was filtered over Celite and washed with methylene chloride. The volatiles were concentrated under reduced pressure and the white residue was redissolved in ethyl acetate. The organic layer was washed with 1 M HCl solution (2×). The aqueous layers were extracted with ethyl acetate (2×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. Purification by flash chromatography (silica, 15% EtOAc/hexanes) affords product as a white foam (5.30 g, 11.8 mmol, 90.1%). $[\alpha]^{20}{}_D$ −12.4 (*c* 0.5, CHCl$_3$); $^1$H NMR (CDCl$_3$, 400 MHz) δ 7.41−7.31 (m, 5H), 7.15 (t, *J* = 1.4 Hz, 1H), 7.04 (t, *J* = 2.0 Hz, 1H), 6.93 (dd, *J* = 1.4, 2.0 Hz, 1H), 5.02 (s, 2H), 4.66 (br s, 1H), 4.32 (d, *J* = 10.4 Hz, 1H), 3.79 (obs. m, 1H), 3.78 (s, 3H), 3.44 (s, 3H), 2.10−2.01 (m, 2H), 1.35 (dt, *J* = 2.6, 12.6, 15.2 Hz, 1H), 0.63 (d, *J* = 6.4 Hz, 3H); $^{13}$C NMR (CDCl$_3$, 100 MHz) δ 171.1, 159.3, 143.6, 136.3, 128.6, 128.1, 127.5, 123.3, 122.6, 117.5, 113.4, 81.7, 75.0, 74.8, 70.2, 56.6, 52.1, 33.3, 29.9, 17.6; IR (neat) $\nu_{max}$ 2952, 2928, 2873, 2826, 1748, 1570, 1441, 1141, 1095, 995, 696 cm$^{-1}$; HRMS (CI, NH$_3$) *m/z* calcd for C$_{22}$H$_{25}$BrO$_5$ [M + 23]$^+$ 471.0783, found 471.0774.

**((2R,3S,5S,6S)-6-(3-(Benzyloxy)-5-bromophenyl)-3-methoxy-5-methyltetrahydro-2*H*-pyran-2-yl)methanol (20b).** To a solution of ester (6.00 g, 13.0 mmol) in diethyl ether (260 mL) at 0 °C was added lithium tetrahydroborate (582 mg, 26.7 mmol). The reaction mixture was stirred for $^1/_2$ h at 0 °C and 1 h at room temperature. Water was carefully added and the reaction mixture was extracted with diethyl ether (3×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. Purification by flash chromatography (silica, 30% EtOAc/hexanes) gives alcohol **20b** as a yellow foam (4.90 g, 11.6 mmol, 89%). $[\alpha]^{20}{}_D$ +16.4 (*c* 0.94, CHCl$_3$); $^1$H NMR (CDCl$_3$, 400 MHz) δ 7.41−7.31 (m, 5H), 7.12 (t, *J* = 1.4 Hz, 1H), 7.04 (t, *J* = 2.0 Hz, 1H), 6.92 (t, *J* = 1.6 Hz, 1H), 5.02 (s, 2H), 4.15 (d, *J* = 8.4 Hz, 1H), 3.94−3.84 (m, 2H), 3.60 (m, 1H), 3.37 (s, 3H), 3.31 (q, *J* = 3.6, 8.0 Hz, 1H), 2.18 (m, 1H), 1.90 (m, 2H), 1.53 (ddd, *J* = 3.6, 10.4, 14.0 Hz, 1H), 0.79 (d, *J* = 6.8 Hz, 3H); $^{13}$C NMR (CDCl$_3$, 75.0 MHz) δ 159.4, 143.8, 136.2, 128.6, 128.1, 127.5, 122.9, 122.7, 117.2, 113.1, 78.8, 75.4, 73.7, 70.2, 60.7, 56.3, 32.2, 30.4, 18.0; IR (neat) $\nu_{max}$ 3433, 2929, 2873, 2825, 1570, 1440, 1270, 1046, 833, 696 cm$^{-1}$; HRMS (CI, NH$_3$) *m/z* calcd for C$_{21}$H$_{25}$BrO$_4$ [M + 23]$^+$ 443.0834, found 443.0838.

**(2R,3S,5R)-6-(3-(Benzyloxy)-5-bromophenyl)-3-methoxy-5-methylhexane-1,2-diol (21b).** To a suspension of aluminum

trichloride (7.75 g, 58.1 mmol) in methylene chloride (500 mL) at −78 °C was added anisole (58.0 mL, 534 mmol) dropwise. The light yellow solution was stirred for an additional 5 min before alcohol **20b** (4.90 g, 11.6 mmol) in methylene chloride (80.0 mL) was added dropwise. The reaction mixture was allowed to warm to 0 °C over 3 h (on its own) and stirred for an additional 1 h at that temperature. The reaction was quenched by slow addition of 0.5 M HCl (80.0 mL). The biphasic mixture was diluted with ammonium chloride and extracted with ethyl acetate (2×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. Purification by flash chromatography (silica, 50% EtOAc/hexanes) yields phenol as a colorless oil (3.60 g, 10.9 mmol, 94.0%).

To a solution of the above phenol (1.35 g, 4.08 mmol) in methylene chloride (24.0 mL) at room temperature was added triethylsilane (5.21 mL, 32.6 mmol) followed by scandium(III) triflate (3.01 g, 6.11 mmol). The sealed tube was closed and the reaction was allowed to run at 42 °C for 48 h. Upon completion, the mixture was cooled to room temperature and the reaction was quenched by addition of water. The aqueous layer was extracted with methylene chloride (3×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. Purification by flash chromatography (silica, 80% EtOAc/hexanes) affords diol as a colorless oil (1.20 g, 3.60 mmol, 88.2%).

To a solution of the above diol (2.90 g, 8.70 mmol) in acetone (50.0 mL) was added potassium carbonate (1.80 g, 13.0 mmol) followed by benzyl bromide (1.14 mL, 9.57 mmol). The heterogeneous mixture was stirred at room temperature for 48 h before it was filtered over Celite and washed with ethyl acetate. The solvent was concentrated under reduced pressure and the resulting residue was purified by flash chromatography (silica, 60% EtOAc/hexanes) to afford diol **21b** as a yellow oil (3.40 g, 8.00 mmol, 92.0%). $[\alpha]^{20}{}_D$ −2.2 (*c* 1.2, CHCl$_3$); $^1$H NMR (CDCl$_3$, 400 MHz) δ 7.41−7.28 (m, 5H), 6.96 (t, *J* = 2.2 Hz, 1H), 6.90 (t, *J* = 1.4 Hz, 1H), 6.69 (t, *J* = 2.2 Hz, 1H), 5.00 (s, 2H), 3.73−3.62 (m, 3H), 3.40 (s, 3H), 3.38 (t, *J* = 3.6 Hz, 1H), 2.58 (A of ABq, *J* = 5.8, 13.2 Hz, 1H), 2.41 (d, *J* = 5.2 Hz, 1H), 2.31 (B of ABq, *J* = 8.8, 13.2 Hz, 1H), 2.11 (dd, *J* = 3.4, 8.0 Hz, 1H), 1.91 (m, 1H), 1.62 (dtd, *J* = 4.2, 9.6, 14.0 Hz, 1H), 1.16 (dtd, *J* = 3.6, 9.6, 14.0 Hz, 1H), 0.85 (d, *J* = 6.4 Hz, 3H); $^{13}$C NMR (CDCl$_3$, 75.0 MHz) δ 159.2, 144.1, 136.4, 128.5, 128.0, 127.4, 124.8, 122.4, 115.3, 114.9, 80.7, 72.7, 70.1, 63.3, 58.4, 43.9, 37.4, 31.1, 19.0; IR (neat) $\nu_{max}$ 3405, 2930, 1603, 1566, 1447, 1269, 1156, 1096, 1028, 851, 736, 697 cm$^{-1}$; HRMS (CI, NH$_3$) *m/z* calcd for C$_{21}$H$_{27}$BrO$_4$ [M + 23]$^+$ 445.0990, found 445.0980.

**(3S,4R,5S,7R)-8-(3-(Benzyloxy)-5-bromophenyl)-5-methoxy-3,7-dimethyloct-1-en-4-ol.** To a solution of diol **21b** (0.270 g, 0.596 mmol) in acetone (6.00 mL) and water (6.00 mL) was added a mixture of sodium bicarbonate (0.150 g, 1.79 mmol) and sodium metaperiodate (0.191 g, 0.893 mmol). The resulting suspension was stirred at room temperature for 2 h. The solution was diluted with water followed by ethyl acetate. The aqueous phase was saturated with solid sodium chloride and extracted with ethyl acetate (3×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. The crude aldehyde **14b** was used in the next step without further purification.

To a flame-dried round-bottomed flask was added 4 Å powdered molecular sieves (600 mesh, 0.330 g), toluene (4.00 mL), and 1.00 M (4*S*,5*S*,*Z*)-diisopropyl 2-(but-2-enyl)-1,3,2-dioxaborolane-4,5-dicarboxylate **13** in toluene (1.66 mL) under an atmosphere of nitrogen. The reaction mixture was allowed to stir for 20 min at room temperature before it was cooled to −78 °C. A solution of aldehyde **14b** (0.200 g, 0.475 mmol) in toluene (1.00 mL) was added dropwise via a cannula. The reaction mixture was stirred at −78 °C for 16 h before it was quenched with 1 N NaOH solution (5 mL). The two-phase

mixture was allowed to warm to room temperature and stirred for 1 h. The solution mixture was filtered over a pad of Celite and the aqueous phase was extracted with diethyl ether (3×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. The residue was purified by flash chromatography (silica, 10% EtOAc/hexanes) to yield homoallylic alcohol as a viscous clear oil, dr 20:1. $[\alpha]^{20}_D$ −18.6 (*c* 1.0, CHCl$_3$); $^1$H NMR (CDCl$_3$, 400 MHz) δ 7.41−7.30 (m, 5H), 6.95 (t, *J* = 2.0 Hz, 1H), 6.91 (t, *J* = 2.0 Hz, 1H), 6.70 (t, *J* = 2.0 Hz, 1H), 5.60 (m, 1H), 5.02 (m, 4H), 3.61 (dt, *J* = 2.4, 9.2 Hz, 1H), 3.33 (s, 3H), 3.25 (dt, *J* = 2.4, 10.8 Hz, 1H), 2.58 (A of ABx, *J* = 5.8, 13.2 Hz, 1H), 2.29 (B of ABx, *J* = 9.0, 13.2 Hz, 1H), 2.22 (obs. m, 1H), 2.05 (d, *J* = 2.4 Hz, 1H), 1.93 (m, 1H), 1.63 (ddd, *J* = 3.2, 10.6, 14.0 Hz, 1H), 1.18 (ddd, *J* = 2.4, 10.4, 14.4 Hz, 1H), 1.12 (d, *J* = 6.4 Hz, 3H), 0.77 (d, *J* = 6.8 Hz, 1H); $^{13}$C NMR (CDCl$_3$, 75.0 MHz) δ 159.2, 144.5, 139.8, 136.4, 128.6, 128.0, 127.5, 124.8, 122.4, 115.3, 115.2, 114.9, 80.0, 73.1, 70.1, 57.0, 44.4, 40.4, 34.4, 30.8, 18.5, 17.5; IR (neat) $\nu_{max}$ 3451, 3067, 2928, 1603, 1567, 1448, 1269, 1087, 916, 697 cm$^{-1}$; HRMS (CI, NH$_3$) *m/z* calcd for C$_{24}$H$_{31}$BrO$_3$ [M + 23]$^+$ 469.1354, found 469.1335.

**1-(Benzyloxy)-3-bromo-5-((2*R*,4*S*,5*R*,6*S*)-4-methoxy-5-(methoxymethoxy)-2,6-dimethyloct-7-enyl)benzene (22b).** To a solution of alcohol (1.80 g, 4.02 mmol) in methylene chloride (20.0 mL) and *N*,*N*-diisopropylethylamine (4.20 mL, 24.1 mmol) at 0 °C was added DMAP (147 mg, 1.21 mmol) and chloromethyl methyl ether (1.22 mL, 16.1 mmol) dropwise. The reaction mixture was allowed to warm to room temperature overnight and was quenched by addition of a saturated solution of sodium bicarbonate. The aqueous layer was extracted with methylene chloride (3×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. The crude product was purified by flash chromatography (silica, 5% EtOAc/hexanes) to yield MOM-ether **22b** as a yellow oil (64%, three steps). $[\alpha]^{20}_D$ +17.2 (*c* 0.9, CHCl$_3$); $^1$H NMR (CDCl$_3$, 400 MHz) δ 7.41−7.30 (m, 5H), 6.94 (t, *J* = 2.0 Hz, 1H), 6.92 (t, *J* = 1.6 Hz, 1H), 6.71 (dd, *J* = 1.6, 2.0 Hz, 1H), 5.67 (m, 1H), 5.00 (m, 4H), 4.81 (A of ABq, *J* = 6.4 Hz, 1H), 4.62 (B of ABq, *J* = 6.8 Hz, 1H), 3.61 (dd, *J* = 2.0, 8.8 Hz, 1H), 3.39 (s, 3H), 3.32 (obst. m, 1H), 3.31 (s, 3H), 2.64 (A of ABx, *J* = 5.2, 13.6 Hz, 1H), 2.27 (obs. q, *J* = 8.8, 15.6 Hz, 1H), 2.24 (obs. B of ABx, *J* = 9.2, 13.6 Hz, 1H), 1.93 (m, 1H), 1.68 (ddd, *J* = 3.2, 10.6, 14.0 Hz, 1H), 1.21 (ddd, *J* = 2.0, 10.2, 14.0 Hz, 1H), 1.11 (d, *J* = 6.8 Hz, 3H), 0.75 (d, *J* = 6.4 Hz, 1H); $^{13}$C NMR (CDCl$_3$, 75.0 MHz) δ 159.2, 144.6, 140.9, 136.5, 128.6, 128.0, 127.5, 124.9, 122.2, 115.2, 114.9, 114.8, 97.4, 80.7, 79.0, 70.0, 57.0, 56.0, 44.3, 40.6, 36.4, 31.0. 18.5, 17.6; IR (neat) $\nu_{max}$ 2957, 2926, 1604, 1448, 1154, 1097, 1032, 918, 696 cm$^{-1}$; HRMS (CI, NH$_3$) *m/z* calcd for C$_{26}$H$_{35}$BrO$_4$ [M + 23]$^+$ 513.1616, found 513.1606.

**1-(Benzyloxy)-3-bromo-5-((2*R*,4*S*,5*R*,6*S*)-4-methoxy-5-(methoxymethoxy)-2,6-dimethyloct-7-ynyl)benzene.** To a solution of alkene **22b** (0.420 g, 0.710 mmol) in a 4:1 mixture of acetone (6.40 mL) and water (1.60 mL) was added *N*-methylmorpholine-*N*-oxide (0.166 g, 1.42 mmol). Osmium tetraoxide (0.20 M in toluene, 530 µL) was added dropwise and the reaction flask was stoppered. The reaction mixture was stirred at room temperature for 4 h before it was quenched with a saturated solution of sodium thiosulfate. The aqueous layer was saturated with solid NaCl and extracted with ethyl acetate (3×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. The crude diol was used without further purification in the next step.

To a solution of crude diol (0.440 g, 0.703 mmol) in dry benzene (14.0 mL) was added potassium carbonate (0.292 g, 2.11 mmol) followed by lead tetraacetate (0.468 g, 1.05 mmol). The reaction mixture was stirred for 30 min before it was quenched with a saturated solution of sodium bicarbonate. The aqueous phase was extracted with ethyl acetate (3×). The

combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. The crude aldehyde **23b** was used directly in the next step.

To a suspension of potassium *tert*-butoxide (0.108 g, 0.967 mmol) in tetrahydrofuran (1.00 mL) under an atmosphere of nitrogen at −78 °C was added dimethyl diazomethylphosphonate (0.166 g, 1.10 mmol) in tetrahydrofuran (1.00 mL) dropwise. The reaction mixture was stirred for 15 min before a −78 °C solution of aldehyde **23b** (0.164 g, 0.276 mmol) in tetrahydrofuran (1.00 mL) was added dropwise. The reaction mixture was stirred for 15 min at −78 °C and was diluted with ethyl acetate and washed with brine. The aqueous layer was extracted with ethyl acetate (3×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. Purification by flash chromatography (silica, 2−5% EtOAc/hexanes) yielded terminal alkyne as a yellow oil (80%). $[\alpha]^{20}_D$ +8.9 (*c* 1.0, CHCl$_3$); $^1$H NMR (CDCl$_3$, 400 MHz) δ 7.41−7.31 (m, 5H), 6.94 (m, 2H), 6.72 (t, *J* = 1.8 Hz, 1H), 5.00 (s, 2H), 4.81 (A of ABq, *J* = 6.8 Hz, 1H), 4.65 (B of ABq, *J* = 6.8 Hz, 1H), 3.72 (dd, *J* = 2.0, 8.8 Hz, 1H), 3.67 (dt, *J* = 2.2, 8.0 Hz, 1H), 3.39 (s, 3H), 3.36 (s, 3H), 2.64 (A of ABx, *J* = 5.4, 13.2 Hz, 1H), 2.51 (m, 1H), 2.28 (B of ABx, *J* = 9.2, 13.6 Hz, 1H), 2.11 (d, *J* = 2.4 Hz, 1H), 1.97 (m, 1H), 1.69 (ddd, *J* = 3.6, 10.2, 14.0 Hz, 1H), 1.30 (d, *J* = 6.8 Hz, 3H), 1.25 (obst. m, 1H), 0.82 (d, *J* = 6.4 Hz, 1H); $^{13}$C NMR (CDCl$_3$, 75.0 MHz) δ 159.3, 144.6, 136.5, 128.6, 128.1, 127.5, 124.9, 122.3, 115.2, 114.9, 97.4, 85.8, 80.7, 78.7, 70.7, 70.1, 57.1, 56.2, 44.2, 36.5, 31.1, 28.2, 18.6, 18.1; IR (neat) $\nu_{max}$ 3297, 2931, 2823, 1567, 1448, 1157, 1097, 1030 920, 697 cm$^{-1}$; HRMS (CI, NH$_3$) *m/z* calcd for C$_{26}$H$_{33}$BrO$_4$ [M + 23]$^+$ 511.1460, found 511.1450.

**1-(Benzyloxy)-3-bromo-5-((2*R*,4*S*,5*R*,6*S*)-4-methoxy-5-(methoxymethoxy)-2,6-dimethylnon-7-ynyl)benzene (11b).** To a solution of terminal alkyne (0.355 g, 0.602 mmol) in tetrahydrofuran (6.00 mL) at −78 °C under an atmosphere of nitrogen was added lithium hexamethyldisilazide (1.0 M in THF, 1.3 mL) dropwise. The reaction mixture was allowed to stir for 30 min at −78 °C before methyl iodide (375 µL, 6.02 mmol) was added dropwise. The reaction mixture was stirred for 1 h at −78 °C and 1 h at room temperature. The reaction was quenched by addition of water. The aqueous phase was extracted with ethyl ether (3×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. Purification by flash chromatography (silica, 4% EtOAc/hexanes) provides methyl alkyne **11b** as a clear oil (334 mg, 0.554 mmol, 92%). $[\alpha]^{20}_D$ +7.9 (*c* 1.7, CHCl$_3$); $^1$H NMR (CDCl$_3$, 400 MHz) δ 7.41−7.30 (m, 5H), 6.94 (m, 2H), 6.73 (br s, 1H), 5.00 (s, 2H), 4.81 (A of ABq, *J* = 6.4 Hz, 1H), 4.64 (B of ABq, *J* = 6.8 Hz, 1H), 3.66 (d, *J* = 8.8 Hz, 1H), 3.38 (s, 3H), 3.35 (s, 3H), 2.66 (A of ABx, *J* = 5.2, 13.6 Hz, 1H), 2.43 (m, 1H), 2.26 (B of ABx, *J* = 9.6, 13.6 Hz, 1H), 1.96 (m, 1H), 1.76 (d, *J* = 2.4 Hz, 3H), 1.67 (ddd, *J* = 3.6, 10.4, 14.4 Hz, 1H), 1.27 (ovlp. m, 1H), 1.24 (d, *J* = 6.8 Hz, 3H), 0.82 (d, *J* = 6.4 Hz, 3H); $^{13}$C NMR (CDCl$_3$, 100 MHz) δ 159.3, 144.7, 136.5, 128.6, 128.0, 127.5, 124.9, 122.3, 115.2, 114.9, 97.3, 80.9, 80.5, 79.2, 78.0, 70.1, 57.0, 56.1, 44.2, 36.5, 31.1, 28.5, 18.6, 18.5, 3.4; IR (neat) $\nu_{max}$ 3065, 3033, 2920, 2822, 1604, 1567, 1448, 1269, 1154, 1033, 920, 697 cm$^{-1}$; HRMS (CI, NH$_3$) *m/z* calcd for C$_{27}$H$_{35}$BrO$_4$ [M + 23]$^+$ 525.1616, found 525.1654.

**(*S*)-5-Oxotetrahydrofuran-2-carboxylic Acid.** To a solution of L-glutamic acid **25** (20.0 g, 136.0 mmol) in water (136 mL) and 2 N HCl (80 mL) at 0 °C was added dropwise a solution of sodium nitrate (11.2 g, 163 mmol) in water (80 mL) over 2 h. After addition was complete, the reaction mixture was allowed to warm to room temperature and stirred for 16 h. The aqueous phase was saturated with solid NaCl and was extracted with ethyl acetate (3×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. The crude yellow solid was used in the next step

without further purification (12.0 g, 92.0 mmol, 80%). $[\alpha]^{20}{}_D$ +13.3 (c 1.1, EtOH); $^1$H NMR (400 MHz, DMSO) $\delta$ 4. 96 (dt, J = 1.2, 5.2 Hz, 2H), 3.35 (m, 1H), 2.16 (m, 2H), −0.16 (br s, 1H); $^{13}$C NMR (75 MHz, CDCl$_3$) 175.8, 174.7, 75.1, 26.6, 25.8; IR (neat) $\nu_{max}$ 2999, 1760, 1419, 1180, 1067 cm$^{-1}$; HRMS (CI, NH$_3$) m/z calcd for C$_5$H$_6$O$_4$ [M+]$^+$ 130.0266, found 130.0250.

**(S)-5-(Hydroxymethyl)dihydrofuran-2(3H)-one (26).** To a solution of (S)-5-oxotetrahydrofuran-2-carboxylic acid (14.0 g, 108.0 mmol) in tetrahydrofuran (100 mL) was added BH$_3$·Me$_2$S (12.3 mL, 130 mmol) over 1 h. The resulting mixture was stirred at room temperature for 4 h. The reaction was quenched with methanol (63 mL) and the resulting mixture was concentrated in vacuo. The residue was purified by flash chromatography (silica, 10% MeOH/CH$_2$Cl$_2$) to afford lactone **26** as a colorless oil (10.0 g, 86.0 mmol, 80%). $[\alpha]^{20}{}_D$ +34.3 (c 1.02, EtOH); $^1$H NMR (400 MHz, CDCl$_3$) $\delta$ 4.62 (m, 1H), 3.89 (dd, J = 2.8, 12.6 Hz, 1H), 3.64 (dd, J = 4.6, 12.6 Hz, 1H), 2.64−2.51 (m, 2H), 2.30−2.12 (m, 3H); $^{13}$C NMR (75 MHz, CDCl$_3$) 177.8, 80.8, 64.0, 28.5, 23.1; IR (neat) $\nu_{max}$ 3423, 2942, 1770, 1356, 1190, 1063, 937 cm$^{-1}$; HRMS (CI, NH$_3$) m/z calcd for C$_5$H$_8$O$_3$ [M+]$^+$ 116.0473, found 116.0451.

**(S)-5-((tert-Butyldimethylsilyloxy)methyl)dihydrofuran-2(3H)-one.** To a solution of alcohol **26** (10.0 g, 86.0 mmol) in N,N-dimethylformamide (45 mL) at 0 °C was added 1H-imidazole (14.6 g, 215 mmol) and tert-butyldimethylsilyl chloride (16.9 g, 112 mmol). The reaction mixture was allowed to warm to room temperature and was stirred for 12 h. Water was added and the aqueous phase was extracted with hexanes (3×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. The residue was purified by flash chromatography (silica, 10% EtOAc/hexanes) to afford a colorless oil (19.8 g, 86.0 mmol, 99%). $[\alpha]^{20}{}_D$ +12.8 (c 1.0, CDCl$_3$); $^1$H NMR (400 MHz, CDCl$_3$) $\delta$ 4.56 (m, 1H), 3.84 (dd, J = 3.2, 11.2 Hz, 1H), 3.66 (dd, J = 2.8, 11.2 Hz, 1H), 2.63−2.55 (m, 1H), 2.48−2.40 (m, 1H), 2.29−2.11 (m, 2H), 0.93 (s, 9H), 0.04 (s, 6H); $^{13}$C NMR (75 MHz, CDCl$_3$) 177.3, 80.1, 64.9, 28.4, 25.7, 23.5, 18.2, −5.5, −5.6; IR (neat) $\nu_{max}$ 2954, 2932, 2858, 1779, 1256, 1122, 838 cm$^{-1}$; HRMS (CI, NH$_3$) m/z calcd for C$_{11}$H$_{22}$O$_3$Si [M + 1]$^+$ 231.1416, found 231.1426.

**(S,E)-Ethyl 7-(tert-Butyldimethylsilyloxy)-6-hydroxy-2-methylhept-2-enoate (27).** To a solution of TBS ether (19.8 g, 86.0 mmol) in toluene (287 mL) at −78 °C was added DibalH (1.0 M in toluene, 93.0 mL, 92.9 mmol). The reaction mixture was stirred for 2 h at that temperature and was quenched by addition of methanol (30 mL). The mixture was then allowed to warm to room temperature and diluted with ethyl acetate, then washed with saturated solutions of NaKtartrate and NaCl. The organic layer was dried over MgSO$_4$, filtered, and concentrated in vacuo. The crude product was used in the next step without further purification.

To a solution of lactol (19.6 g, 86.0 mmol) in benzene (453 mL) was added added (carbethoxyethylidene)triphenyl phosphorane (37.4 g, 103 mmol). The reaction mixture was allowed to reflux for 2 h. The mixture was cooled to room temperature and concentrated in vacuo. The residue was purified by flash chromatography (silica, 15% EtOAc/hexanes) to provide alcohol **27** as a colorless oil (19.8 g, 62.6 mmol, 73%). $[\alpha]^{20}{}_D$ +2.6 (c 0.99, CDCl$_3$); $^1$H NMR (400 MHz, CDCl$_3$) $\delta$ 6.73 (t, J = 6.8 Hz, 1H), 4.16 (q, J = 6.8, 14.4 Hz, 2H), 3.61 (m, 2H), 3.38 (t, J = 8.4 Hz, 1H), 2.31 (m, 2H), 1.83 (s, 3H), 1.55 (m, 2H), 1.28 (t, J = 7.0 Hz, 3H), 0.88 (s, 9H), 0.05 (s, 6H); $^{13}$C NMR (75 MHz, CDCl$_3$) 168.1, 141.4, 128.3, 71.2, 67.1, 60.3, 31.7, 25.9, 24.8, 18.3, 14.2, 12.3, −5.4; IR (neat) $\nu_{max}$ 3501, 2954, 2930, 2858, 1711, 1258, 1096, 838 cm$^{-1}$; HRMS (CI, NH$_3$) m/z calcd for C$_{16}$H$_{32}$O$_4$Si [M + 1]$^+$ 317.2148, found 317.2156.

**(S,E)-Ethyl 7-(tert-Butyldimethylsilyloxy)-6-methoxy-2-methylhept-2-enoate.** To a solution of alcohol **27** (18.0 g, 56.9 mmol) in dichloromethane (1140 mL) was added 4 Å powdered molecular sieves (600 mesh, 37.0 g), Proton-Sponge (37.0 g, 171 mmol), and Me$_3$OBF$_4$ (21.0 g, 142 mmol). The resulting mixture was stirred at room temperature for 3 h, filtered over Celite, and concentrated in vacuo. The residue was dissolved in ethyl acetate and washed with 1 M CuSO$_4$ and NaCl solutions. The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. The residue was purified by flash chromatography (silica, 7% EtOAc/hexanes) to afford a colorless oil (17.8 g, 53.9 mmol, 95%). $[\alpha]^{20}{}_D$ −9.0 (c 0.95, CHCl$_3$); $^1$H NMR (400 MHz, CDCl$_3$) $\delta$ 6.75 (t, J = 7.2 Hz, 1H), 4.17 (q, J = 7.0, 14.3 Hz, 2H), 3.63 (dd, J = 5.5, 10.4 Hz, 1H), 3.53 (dd, J = 5.5, 10.4 Hz, 1H), 3.39 (s, 3H), 3.17 (m, 1H), 2.25 (q, J = 7.9, 15.2 Hz, 2H), 1.82 (s, 3H), 1.68- 1.51 (m, 2H), 1.27 (t, J = 7.0 Hz, 3H), 0.87 (s, 9H), 0.04 (s, 6H); $^{13}$C NMR (75 MHz, CDCl$_3$) 168.1, 141.8, 128.1, 81.1, 64.8, 60.3, 57.8, 30.2, 25.9, 24.5, 18.2, 14.2, 12.2, −5.4; IR (neat) $\nu_{max}$ 2954, 2931, 2858, 1713, 1257, 1117, 838 cm$^{-1}$; HRMS (CI, NH$_3$) m/z calcd for C$_{17}$H$_{34}$O$_4$Si [M + 1]$^+$ 331.2305, found 331.2287.

**(S,E)-Ethyl 7-Hydroxy-6-methoxy-2-methylhept-2-enoate (28).** To TBS ether (17.8 g, 53.9 mmol) was added a 2% solution of HCl in methanol (900 mL). The reaction mixture was stirred for 1 h at room temperature and concentrated in vacuo. The residue was purified by flash chromatography (silica, 30% EtOAc/hexanes) to afford primary alcohol **28** as a colorless oil (11.5 g, 53.2 mmol, 99%). $[\alpha]^{20}{}_D$ +10.8 (c 1.01, CHCl$_3$); $^1$H NMR (400 MHz, CDCl$_3$) $\delta$ 6.72 (dt, J = 1.5, 7.6 Hz, 1H), 4.17 (q, J = 7.3, 14.0 Hz, 2H), 3.70 (dd, J = 3.7, 11.6 Hz, 1H), 3.49 (dd, J = 5.5, 11.6 Hz, 1H), 3.39 (s, 3H), 3.26 (m, 1H), 2.22 (q, J = 7.3, 15.9 Hz, 2H), 1.82 (s, 3H), 1.71 (m, 1H), 1.59 (m, 1H), 1.27 (t, J = 7.0 Hz, 3H); $^{13}$C NMR (75 MHz, CDCl$_3$) 167.9, 141.1, 128.2, 80.8, 63.4, 60.3, 57.0, 29.4, 24.3, 14.1, 12.1; IR (neat) $\nu_{max}$ 3448, 2980, 2933, 1709, 1273, 1095, 746 cm$^{-1}$; HRMS (CI, NH$_3$) m/z calcd for C$_{11}$H$_{20}$O$_4$ [M + 1]$^+$ 217.1440, found 217.1422.

**(S,E)-Ethyl 6-Methoxy-2-methyl-7-oxohept-2-enoate (12a).** To oxalyl chloride (156 µL, 1.85 mmol) in methylene chloride (2.40 mL) at −78 °C was added dimethyl sulfoxide (262 µL, 3.70 mmol) dropwise. The reaction mixture was stirred for 15 min before alcohol **28** (200 mg, 0.925 mmol) in methylene chloride (5.00 mL) was added via a cannula. The mixture was stirred for $^1/_2$ h before triethylamine (0.773 mL, 5.55 mmol) was added dropwise. The reaction was allowed to warm to room temperature for 4 h and water was added. The aqueous phase was extracted with dichloromethane (3×). The combined organic layers were washed with brine, dried over MgSO$_4$, filtered, and concentrated. The residue was purified by flash chromatography (silica, 10% EtOAc/hexanes) to afford aldehyde **12a** as a colorless oil (197 mg, 0.919 mmol, 99%). $[\alpha]^{20}{}_D$ −52.1 (c 1.75, CDCl$_3$); $^1$H NMR (400 MHz, CDCl$_3$) $\delta$ 9.62 (d, J = 2.0 Hz, 1H), 6.65 (dt, J = 1.2, 7.2 Hz, 1H), 4.13 (q, J = 7.4 14.2 Hz, 2H), 3.51 (m, 1H), 3.40 (s, 3H), 2.25 (m, 2H), 1.79 (s, 3H), 1.74 (m, 2H), 1.24 (t, J = 7.2 Hz, 3H); $^{13}$C NMR (75 MHz, CDCl$_3$) 203.5, 168.0, 139.9, 128.5, 84.7, 60.5, 58.3, 28.5, 23.7, 14.2, 12.3; IR (neat) $\nu_{max}$ 2984, 2828, 1734, 1709, 1266, 1133 cm$^{-1}$; HRMS (CI, NH$_3$) m/z calcd for C$_{11}$H$_{18}$O$_4$ [M+]$^+$ 214.1205, found 214.1203.

**(2E,6S,7S,8E,10S,11R,12S)-Ethyl 15-(3-(Benzyloxy)-5-bromophenyl)-7-hydroxy-6,12-dimethoxy-11-(methoxymethyl)-2,8,10-trimethylpentadeca-2,8-dienoate (33b).** To a sealed tube containing **11b** (15.0 mg, 0.0299 mmol) was added toluene (300 µL). Bis(cyclopentadienyl)zirconium chloride hydride (15.2 mg, 0.0600 mmol) was added and the reaction mixture was heated to 50 °C. After 6 h, the reaction tube was allowed to cool to room temperature and then it was cooled to −65 °C. A solution of 2.00 M dimethylzinc in toluene (15.4 µL) was added dropwise and the solution mixture was warmed to 0 °C. Aldehyde **12a** (7.83 mg, 0.0366 mmol) was added dropwise to the reaction mixture dropwise. After 1 h at 0 °C, the reaction mixture was quenched with a saturated solution of ammonium chloride. The aqueous layer was extracted with diethyl ether (3×). The combined organic layers were dried with magnesium sulfate, filtered,

and concentrated under reduced pressure. Purification by flash chromatography (silica, 30% EtOAc/hexanes) gives reproducible 40−89% yields of desired product **33b** as a yellow oil. $[\alpha]^{20}_D$ +12.2 (*c* 1.1, CHCl$_3$); $^1$H NMR (CDCl$_3$, 400 MHz) $\delta$ 7.43−7.28 (m, 5H), 6.94 (s, 1H), 6.91 (s, 1H), 6.71 (s, 1H), 6.66 (t, *J* = 7.4 Hz, 1H), 5.29 (d, *J* = 10.0 Hz, 1H), 4.99 (s, 2H), 4.81 (A of ABq, *J* = 6.4 Hz, 1H), 4.60 (B of ABq, *J* = 6.4 Hz, 1H), 4.13 (q, *J* = 7.4, 14.6 Hz, 1H), 3.84 (dd, *J* = 3.2, 6.4 Hz, 1H), 3.60 (d, *J* = 9.2 Hz, 1H), 3.40 (s, 3H), 3.38 (s, 3H), 3.28 (s, 3H), 3.20 (m, 2H), 2.64 (dd, *J* = 5.2, 13.6 Hz, 1H), 2.54 (d, *J* = 2.8 Hz, 1H), 2.51 (m, 1H), 2.24 (m, 3H), 1.94 (m, 1H), 1.81 (s, 3H), 1.72 (m, 1H), 1.61 (s, 3H), 1.54 (m, 1H), 1.28 (ovlp. m, 1H), 1.23 (t, *J* = 6.4 Hz, 1H), 1.14 (m, 1H), 1.06 (d, *J* = 6.4 Hz, 1H), 0.72 (d, *J* = 6.4 Hz, 1H); $^{13}$C NMR (CDCl$_3$, 75.0 MHz) $\delta$ 167.8, 159.2, 144.3, 140.9, 136.4, 133.8, 131.4, 128.5, 128.2, 128.0, 127.4, 124.8, 122.3, 115.1, 114.9, 97.3, 81.2, 81.0, 79.4, 78.6, 70.0, 60.3, 58.4, 56.8, 56.0, 44.1, 36.4, 34.4, 30.8, 29.3, 23.9, 18.6, 17.7, 14.2, 12.3; IR (neat) $\nu_{max}$ 3457, 2928, 1708, 1448, 1269, 1097, 1030, 697 cm$^{-1}$; HRMS (CI, NH$_3$) *m/z* calcd for C$_{38}$H$_{55}$BrO$_8$ [M + 23]$^+$ 741.2978, found 741.3000.

**(2*E*,6*S*,7*S*,8*E*,10*S*,12*S*,14*R*)-Ethyl 15-(3-(Benzyloxy)-5-bromophenyl)-7-*tert*-butyldimethylsilyloxy)-6,12-dimethoxy-11-(methoxymethoxy)-2,8,10,14-tetramethylpentadeca-2,8-dienoate.** To a solution of alcohol **33b** (13.0 mg, 0.0173 mmol) in methylene chloride (200 μL) at 0 °C was added 2,6-lutidine (8.03 μL, 0.0694 mmol) and *tert*-butyldimethylsilyl trifluoromethanesulfonate (7.96 μL, 0.0347 mmol). The reaction mixture was allowed to stir for 2 h at 0 °C and was quenched by addition of a saturated solution of sodium bicarbonate. The aqueous layer was extracted with methylene chloride (3×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. Purification by flash chromatography (silica, 20% EtOAc/hexanes) provided 95% of the desired product as a yellow oil. $[\alpha]^{20}_D$ +16.9 (*c* 1.3, CHCl$_3$); $^1$H NMR (CDCl$_3$, 400 MHz) $\delta$ 7.41−7.29 (m, 5H), 6.93 (s, 1H), 6.91 (s, 1H), 6.71 (s, 1H), 6.64 (dt, *J* = 1.6, 7.4 Hz, 1H), 5.13 (d, *J* = 10.0 Hz, 1H), 4.99 (s, 1H), 4.83 (A of ABq, *J* = 6.4 Hz, 1H), 4.60 (B of ABq, *J* = 6.4 Hz, 1H), 4.11 (q, *J* = 7.0, 14.2 Hz, 1H), 3.94 (d, *J* = 6.8 Hz, 1H), 3.59 (d, *J* = 9.6 Hz, 1H), 3.44 (s, 3H), 3.38 (s, 3H), 3.26 (s, 3H), 3.18 (d, *J* = 10.8 Hz, 1H), 3.06 (ddd, *J* = 2.8, 7.0, 10.0 Hz, 1H), 2.64 (dd, *J* = 5.2, 13.6 Hz, 1H), 2.46 (m, 1H), 2.23 (m, 3H), 1.93 (m, 1H), 1.79 (s, 3H), 1.73 (m, 1H), 1.56 (s, 3H), 1.46 (m, 1H), 1.30 (m, 1H), 1.21 (t, *J* = 7.2 Hz, 3H), 1.12 (m, 1H), 1.04 (d, *J* = 6.8 Hz, 3H), 0.87 (s, 9H), 0.71 (d, *J* = 6.4 Hz, 1H), 0.05 (s, 3H), −0.01 (s, 3H); $^{13}$C NMR (CDCl$_3$, 75.0 MHz) $\delta$ 168.0, 159.3, 144.5, 141.5, 136.5, 135.0, 130.5, 128.6, 128.0, 127.5, 124.9, 122.3, 115.2, 114.9, 97.4, 83.4, 81.0, 80.6, 79.2, 70.1, 60.3, 59.8, 56.8, 56.1, 44.2, 36.4, 30.8, 30.1, 25.8, 24.9, 18.7, 18.1, 17.8, 14.2, 12.5, 12.4, −4.7, −4.9; IR (neat) $\nu_{max}$ 2955, 2928, 2856, 2823, 1710, 1448, 1258, 1111, 1031, 837, 776 cm$^{-1}$; HRMS (CI, NH$_3$) *m/z* calcd for C$_{44}$H$_{69}$BrO$_8$Si [M + 23]$^+$ 855.3843, found 855.3819.

**(2*E*,6*S*,7*S*,8*E*,10*S*,12*S*,14*R*)-15-(3-(Benzyloxy)-5-bromophenyl)-7-*tert*-butyldimethylsilyloxy)-6,12-dimethoxy-11-(methoxymethoxy)-2,8,10,14-tetramethylpentadeca-2,8-dienamide.** Into a 10 mL round-bottomed flask containing ester (12.0 mg, 0.0139 mmol) was added a mixture of tetrahydrofuran (400 μL), methanol (400 μL), and water (200 μL). Lithium hydroxide, monohydrate (11.6 mg, 0.278 mmol) was added and reaction mixture was allowed to stir for 48 h. The reaction mixture was concentrated under reduced pressure. The residue was diluted with pH 4.5 NaH$_2$PO$_4$ (10 mL) and extracted with methylene chloride (5×) with the aqueous phase saturated with solid NaCl each time. The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. The crude acid was used in the next step without further purification.

To a solution of acid (17.0 mg, 0.0203 mmol) in methylene chloride (1.00 mL) at −20 °C was added sequentially triethylamine (85.4 μL, 0.50 M solution in methylene chloride) and ethyl

chloroformate (46.8 μL, 0.50 M solution in methylene chloride). The reaction mixture was allowed to stir for 30 min at −20 °C (checked by TLC) before anhydrous ammonia (*l*) was bubbled into the solution (until the disappearance of the mixed anhydride by TLC). The reaction was quenched with water and warmed to room temperature. The aqueous layer was extracted with methylene chloride (3×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. Purification by flash chromatography (silica, 50% EtOAc/hexanes) provided 60% (2 steps) of the desired product as a yellow oil and recovered acid in 40%. $[\alpha]^{20}_D$ +10.5 (*c* 1.0, CHCl$_3$); $^1$H NMR (CDCl$_3$, 400 MHz) $\delta$ 7.41−7.29 (m, 5H), 6.94 (s, 1H), 6.91 (s, 1H), 6.71 (s, 1H), 6.32 (t, *J* = 6.6 Hz, 1H), 5.41 (br s, 2H), 5.13 (d, *J* = 9.6 Hz, 1H), 4.99 (s, 2H), 4.82 (A of ABq, *J* = 6.8 Hz, 1H), 4.60 (B of ABq, *J* = 6.4 Hz, 1H), 3.94 (d, *J* = 6.8 Hz, 1H), 3.59 (d, *J* = 9.6 Hz, 1H), 3.43 (s, 3H), 3.38 (s, 3H), 3.27 (s, 3H), 3.18 (d, *J* = 10.8 Hz, 1H), 3.07 (ddd, *J* = 2.4, 6.8, 9.6 Hz, 1H), 2.65 (dd, *J* = 5.2, 13.6 Hz, 1H), 2.46 (m, 1H), 2.21 (m, 3H), 1.92 (m, 1H), 1.80 (s, 3H), 1.71 (ddd, *J* = 3.6, 10.8, 14.4 Hz, 1H), 1.56 (s, 3H), 1.46 (m, 1H), 1.27 (m, 1H), 1.13 (m, 1H), 1.03 (d, *J* = 6.4 Hz, 3H), 0.87 (s, 9H), 0.71 (d, *J* = 6.4 Hz, 1H), 0.05 (s, 3H), −0.01 (s, 3H); $^{13}$C NMR (CDCl$_3$, 100 MHz) $\delta$ 171.2, 159.2, 144.5, 137.3, 136.4, 134.9, 130.5, 130.0, 128.6, 128.1, 127.5, 124.9, 122.3, 115.2, 115.0, 97.4, 83.6, 81.1, 80.3, 79.4, 70.1, 59.7, 56.8, 56.1, 44.2, 36.4, 34.3, 30.9, 30.2, 25.8, 24.8, 18.8, 18.1, 17.6, 12.64, 12.56, −4.7, −4.9; IR (neat) $\nu_{max}$ 3342, 2954, 2927, 1684, 1457, 1376, 1250, 1110, 1030, 837 cm$^{-1}$; HRMS (CI, NH$_3$) *m/z* calcd for C$_{42}$H$_{66}$BrNO$_7$Si [M + 23]$^+$ 826.3690, found 826.3662.

**(4*E*,8*S*,9*S*,10*E*,12*S*,13*R*,14*S*,16*R*)-20-(Benzyloxy)-9-(*tert*-butyldimethylsilyloxy)-8,14-dimethoxy-13-(methoxymethoxy)-4,10,12,16-tetramethyl-2-aza-bicyclo[16.3.1]docosa-1(21),4,10,-18(22),19-pentaen-3-one (34b).** To a solution of amide (10.0 mg, 0.0120 mmol) in toluene (0.800 mL) in a sealed tube was added potassium carbonate (4.96 mg, 0.0359 mmol). Copper(I) iodide (1.1 mg, 0.0060 mmol) and *N*,*N*′-dimethyl-1,2-ethanediamine (1.28 μL, 0.0120 mmol) were added sequentially and the tube was sealed closed. The green suspension was heated to 100 °C for 36 h. The contents were filtered over plug of silica and washed with ethyl acetate. Concentration of the solvent followed by flash chromatography (silica, 50% EtOAc/hexanes) yields product **34b** in 82% yield. $[\alpha]^{20}_D$ +49.6 (*c* 1.3, CHCl$_3$); $^1$H NMR (CDCl$_3$, 400 MHz) $\delta$ 7.41−7.30 (m, 5H), 7.13 (br s, 1H), 6.69 (br s, 1H), 6.55 (s, 1H), 6.36 (s, 1H), 5.85 (t, *J* = 6.0 Hz, 1H), 5.00 (s, 2H), 4.98 (obst. d, 1H), 4.81 (A of ABq, *J* = 6.4 Hz, 1H), 4.65 (B of ABq, *J* = 6.8 Hz, 1H), 3.77 (d, *J* = 7.6 Hz, 1H), 3.54 (d, *J* = 8.8 Hz, 1H), 3.45 (s, 3H), 3.41 (s, 3H), 3.31 (s, 3H), 3.12 (d, *J* = 10.0 Hz, 1H), 2.98 (dt, *J* = 3.0, 7.8 Hz, 1H), 2.78 (dd, *J* = 4.0, 13.2 Hz, 1H), 2.43 (m, 1H), 2.36 (dd, *J* = 6.4, 13.6 Hz, 1H), 2.20 (m, 1H), 2.11 (m, 1H), 1.91 (m, 1H), 1.74 (s, 3H), 1.62 (m, 1H), 1.53 (s, 1H), 1.25 (m, 1H), 1.17−1.08 (m, 2H), 1.04 (d, *J* = 6.8 Hz, 1H), 0.86 (s, 9H), 0.72 (d, *J* = 6.8 Hz, 1H), 0.04 (s, 3H), −0.03 (s, 3H); $^{13}$C NMR (CDCl$_3$, 75.0 MHz) $\delta$ 173.0, 159.1, 142.1, 139.1, 136.6, 136.1, 134.6, 131.1, 131.0, 128.6, 128.0, 127.5, 118.0, 113.8, 105.7, 97.6, 83.3, 82.4, 82.0, 79.6, 70.0, 60.8, 57.0, 56.2, 42.9, 34.4, 33.5, 31.3, 30.7, 25.8, 23.7, 18.4, 18.1, 17.6, 13.7, 11.0, −4.7, −4.9; IR (neat) $\nu_{max}$ 3300, 2927, 2856, 1664, 1594, 1461, 1111, 1032, 837 cm$^{-1}$; HRMS (CI, NH$_3$) *m/z* calcd for C$_{42}$H$_{65}$NO$_7$Si [M + 23]$^+$ 746.4428, found 746.4434.

**(4*E*,8*S*,9*S*,10*E*,12*S*,13*R*,14*S*,16*R*)-Carbamic Acid 20-(Benzyloxy)-8,14-dimethoxy-13-(methoxymethoxy)-4,10,12,16-tetramethyl-3-oxo-2-aza-bicyclo[16.3.1]docosa-1(21),4,10,18(22),19-pentaen-9-yl Ester.** To a solution of TBS ether **34b** (13.0 mg, 0.0172 mmol) in tetrahydrofuran (1.30 mL) in a nalgene vial was added a premixed solution of [pyridine hydrofluoride:pyridine: THF (1:1:2.5) by volume] (600 μL) dropwise. The reaction was allowed to run at room temperature for 24 h before another 600 μL of HF mixture was added. The reaction was stirred for

another 12 h and then was quenched by addition of sodium bicarbonate. The aqueous layer was extracted with ethyl acetate (3×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. The crude product was purified by flash chromatography (silica, 50−100% EtOAc/hexanes) to yield secondary alcohol as a clear glass. This material was used immediately in the next step without further purification.

To a solution of the above alcohol (4.0 mg, 0.0063 mmol) in $CH_2Cl_2$ (800 $\mu$L) was added trichloroacetyl isocyanate (1.5 $\mu$L, 0.0126 mmol). The reaction was stirred for 15 min and methanol (1.0 mL) was added followed by potassium carbonate (4.0 mg). The reaction was stirred for $^1/_2$ h (monitor by TLC) before solvent was evaporated in vacuo. Purification by flash chromatography (silica, 80% EtOAc/hexanes) provided 80% (2 steps) of the desired product as a clear glass. $[\alpha]^{20}_D$ +49.3 (*c* 0.8, CHCl$_3$); $^1$H NMR (CDCl$_3$, 400 MHz) $\delta$ 8.08 (br s, 1H), 7.43−7.28 (m, 6H), 6.53 (s, 1H), 6.39 (s, 1H), 5.98 (t, $J$ = 6.4 Hz, 1H), 5.35 (d, $J$ = 10.0 Hz, 1H), 5.17 (d, $J$ = 4.0 Hz, 1H), 5.02 (s, 2H), 4.83 (A of ABq, $J$ = 6.4 Hz, 1H), 4.78 (br s, 2H), 4.63 (B of ABq, $J$ = 6.4 Hz, 1H), 3.59 (d, $J$ = 9.2 Hz, 1H), 3.41 (s, 3H), 3.38 (s, 3H), 3.34 (m, 1H), 3.29 (s, 3H), 3.20 (m, 1H), 2.74 (d, $J$ = 10.0 Hz, 1H), 2.52−2.38 (m, 2H), 2.18 (m, 2H), 1.83 (s, 3H), 1.61 (m, 2H), 1.52 (s, 3H), 1.44 (m, 2H), 1.28 (m,1H), 1.07 (d, $J$ = 6.8 Hz, 3H), 0.79 (d, $J$ = 6.4 Hz, 3H); $^{13}$C NMR (CDCl$_3$, 75.0 MHz) $\delta$ 171.2, 159.1, 156.5, 143.4, 139.5, 136.8, 133.9, 131.1, 129.7, 128.5, 127.9, 127.6, 115.4, 113.1, 104.4, 97.3, 83.5, 80.6, 80.1, 79.9, 77.2, 70.0, 59.0, 56.6, 56.1, 42.9, 42.2, 35.5, 34.5, 29.4, 25.0, 19.7, 17.8, 13.3, 13.0; IR (neat) $\nu_{max}$: 3345, 2927, 1733, 1653, 1457, 1375, 1154, 1106, 1032, 837, 740 cm$^{-1}$; HRMS (CI, NH$_3$) $m/z$ calcd for $C_{37}H_{52}N_2O_8$ [M + 23]$^+$ 675.3621, found 675.3606.

**Autolytimycin (2).** To a suspension of aluminum trichloride (11.7 mg, 0.0879 mmol) in methylene chloride (2.20 mL) at −78 °C was added anisole (2.90 mL) dropwise. The light yellow solution was stirred for an additional 5 min before carbamate (6.0 mg, 0.00879 mmol) in methylene chloride (0.700 mL) was added dropwise. The reaction mixture was allowed to warm to 0 °C over 3 h (on its own) and stirred for an additional 1 h at room temperature. The reaction was quenched by slow addition of 0.5 M HCl. The biphasic mixture was diluted with ammonium chloride and extracted with ethyl acetate (3×). The combined organic layers were dried with magnesium sulfate, filtered, and concentrated under reduced pressure. Purification by flash chromatography (silica, 1−5% MeOH/CH$_2$Cl$_2$) yields autolytimycin (**2**) as a white solid in 76% yield. $[\alpha]^{20}_D$ +32.5 (*c* 0.4, MeOH). Mp 246 °C; $^1$H NMR (*d*$_6$-DMSO, 50 °C, 300 MHz) $\delta$ 9.20 (s, 1H), 9.13 (s, 1H), 6.65 (s, 1H), 6.34 (br s, 2H), 6.25 (s, 2H), 5.75 (br t, $J$ = 6.2 Hz, 1H), 5.25 (d, $J$ = 9.6 Hz, 1H), 4.86 (d, $J$ = 7.2 Hz, 1H), 4.18 (d, $J$ = 4.8 Hz, 1H), 3.38 (m, 1H), 3.32 (s, 3H), 3.24 (obst. m, 1H), 3.23 (s, 3H), 3.01 (m, 1H), 2.54 (dd, $J$ = 5.1, 13.2 Hz, 1H), 2.35 (m, 1H), 2.28 (dd, $J$ = 6.0, 12.9, Hz, 1H), 2.17 (m, 1H), 2.07 (m, 1H), 1.78 (m, 1H), 1.71 (s, 3H), 1.51 (m, 1H), 1.41 (s, 3H), 1.29 (m, 1H), 1.25 (m, 1H), 1.17 (m, 1H), 0.92 (d, $J$ = 6.6 Hz, 3H), 0.75 (d, $J$ = 6.9 Hz, 1H); $^{13}$C NMR (*d*$_6$-DMSO, 50 °C, 75 MHz) $\delta$ 170.8, 157.4, 156.1, 141.2, 140.1, 134.2, 133.1, 131.8, 129.9, 115.0, 112.6, 105.8, 80.7, 80.65, 79.5, 73.4, 58.2, 56.3, 42.7, 34.0, 33.5, 30.5, 29.6, 23.3, 18.7, 16.6, 13.2, 11.7; IR (neat) $\nu_{max}$ 3332, 3272, 3197, 2913, 2877, 2824, 1711, 1653, 1617, 1593, 1399, 1383, 1109, 1039, 872 cm$^{-1}$; HRMS (CI, NH$_3$) $m/z$ calcd for $C_{28}H_{42}N_2O_7$ [M + 23]$^+$ 541.2890, found 541.2900.

**Supporting Information Available:** Experimental details and selected spectral data for all new compounds. This material is available free of charge via the Internet at http://pubs.acs.org.